

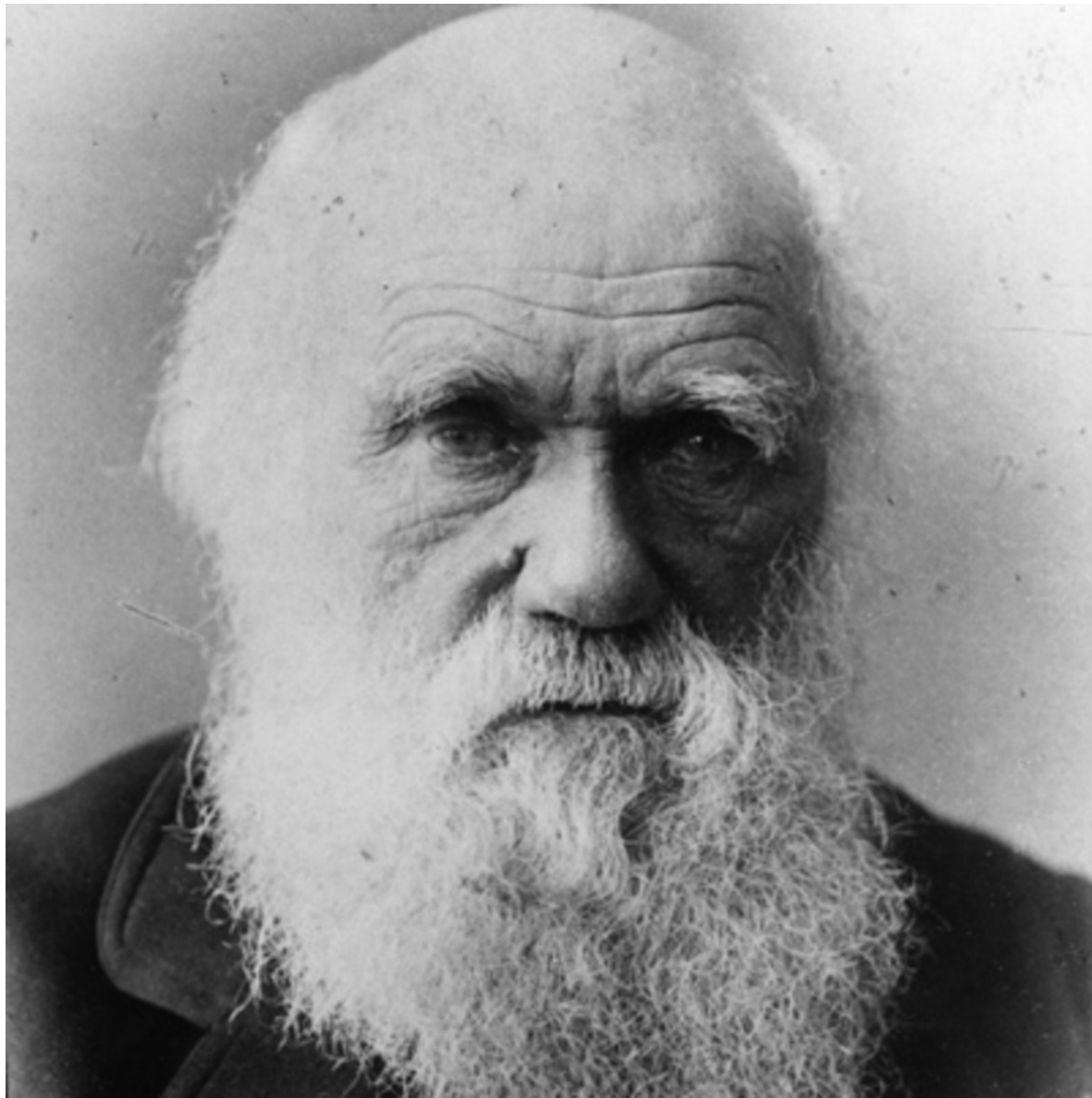
Utveckling av nya molekylärbiologiska metoder

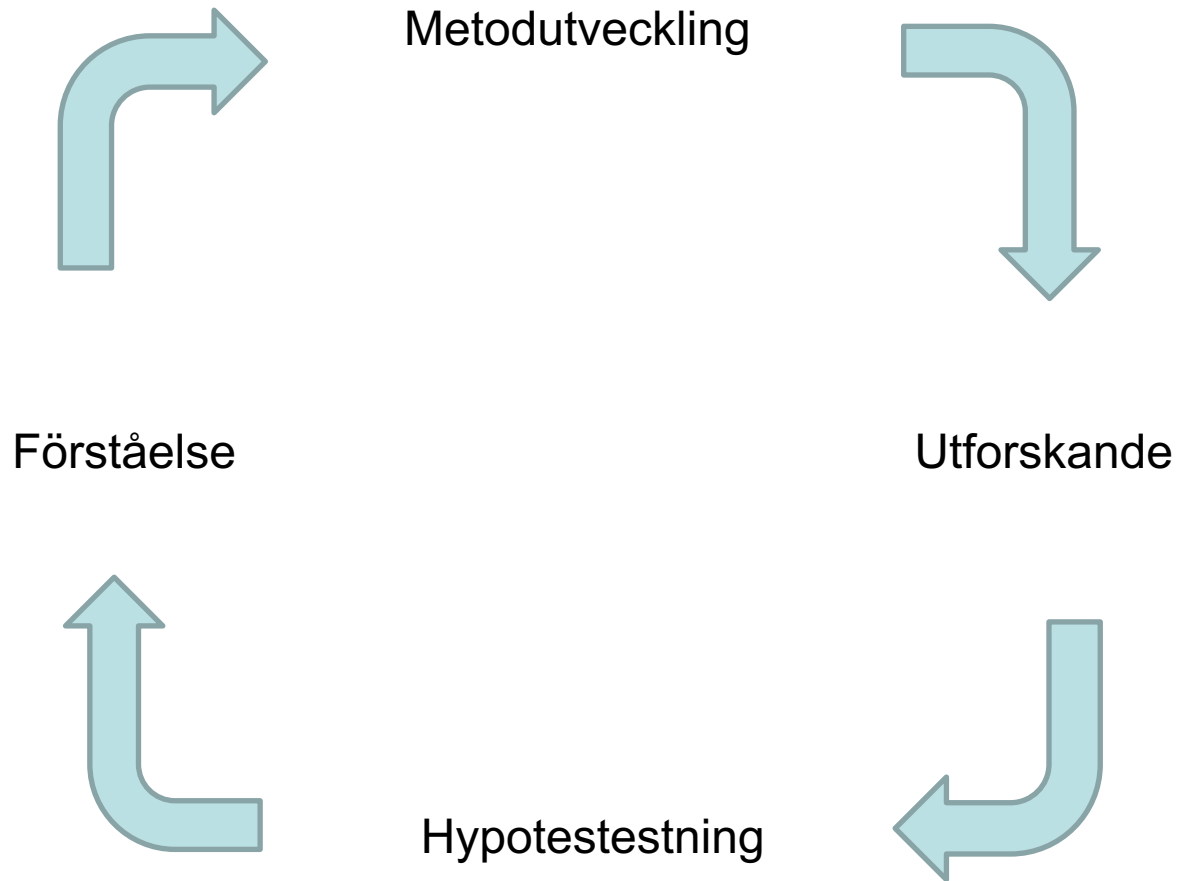
Ola Söderberg, Uppsala University

Vad kan en apotekare arbeta med?

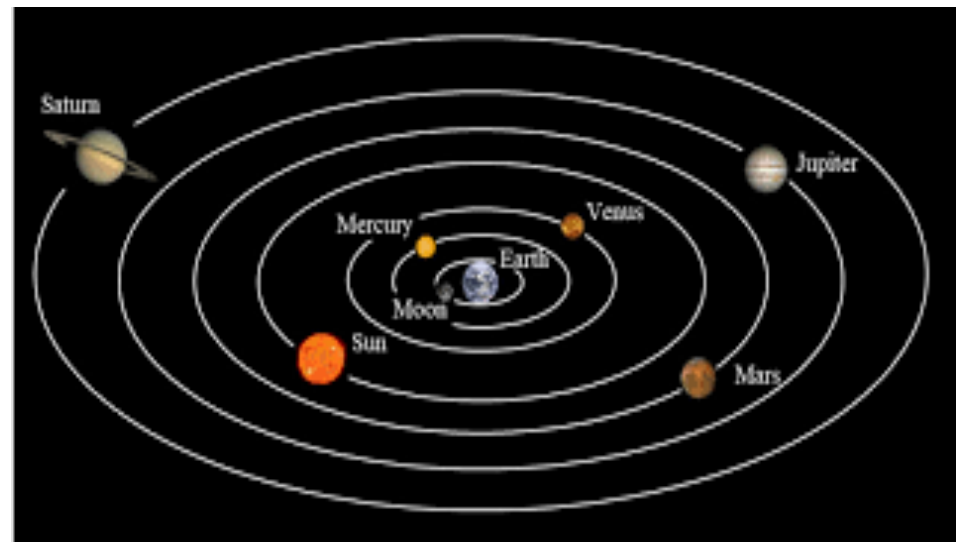
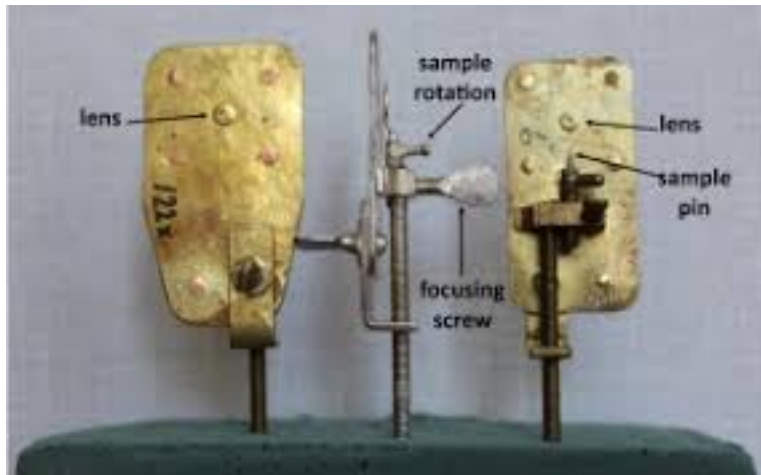
Vad kan ni använda allt ni lärt er till?

Vad är forskning?



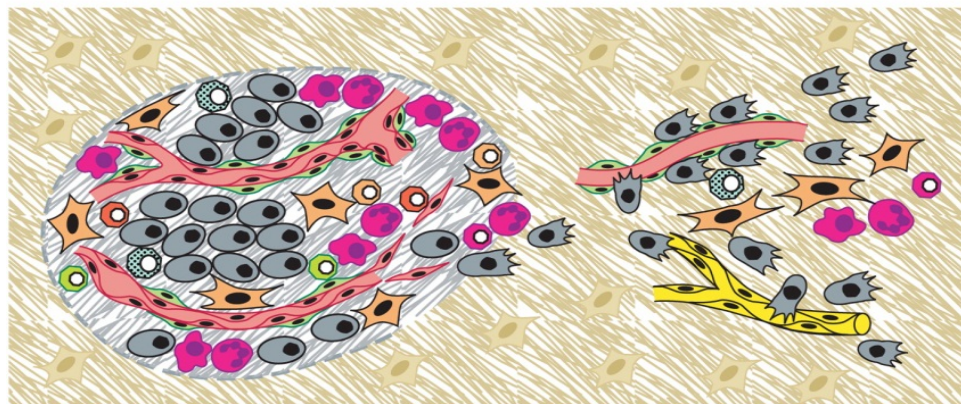
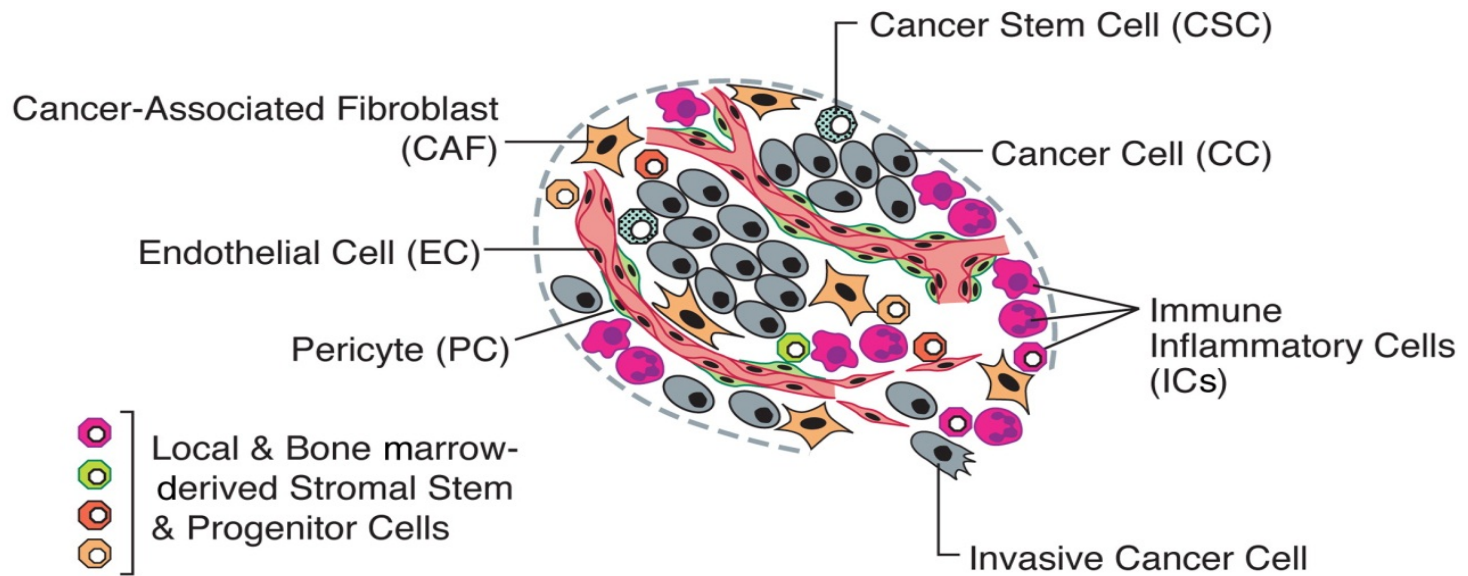


Metodutveckling ger nya verktyg och nya möjligheter

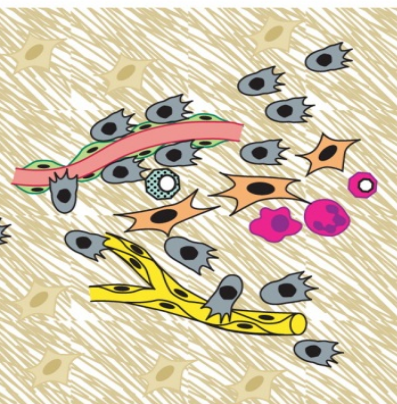


Vad är problemet vi vill lösa?

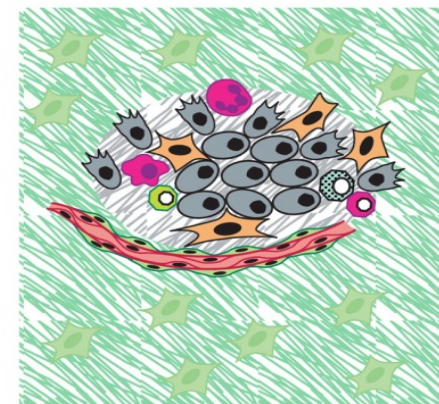
Vad skulle vi vilja studera?



Core of Primary Tumor microenvironment



Invasive Tumor microenvironment



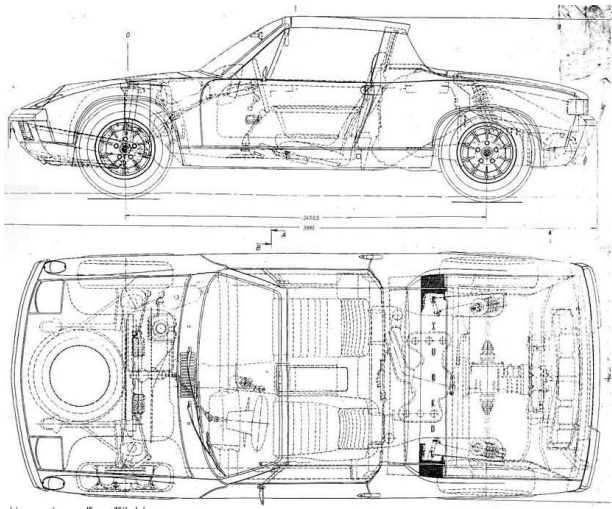
Metastatic Tumor microenvironment





RNA

DNA



Proteins



Protein complexes



Hur kan man göra en ny metod?

Vi använder komponenter från
naturvetenskapen





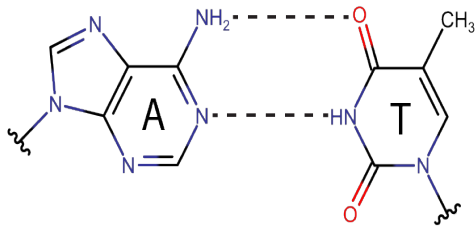


DNA är en enkel och förutsägbar
molekyl

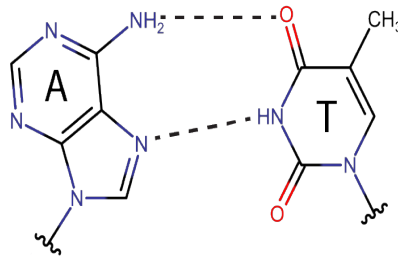
En stor repertoir av DNA-
modifierande enzym

DNA base pairing

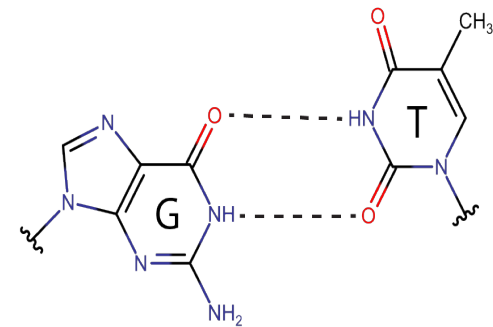
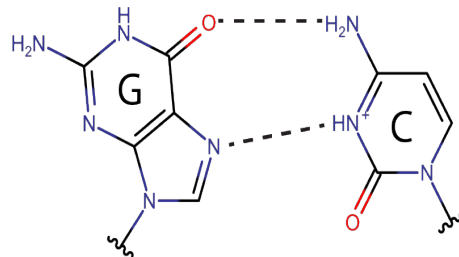
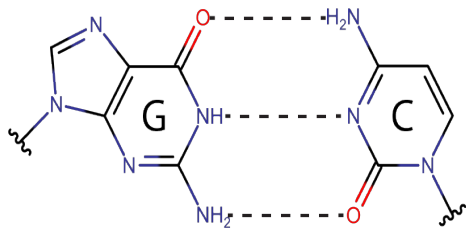
A Watson & Crick



B Hoogsteen

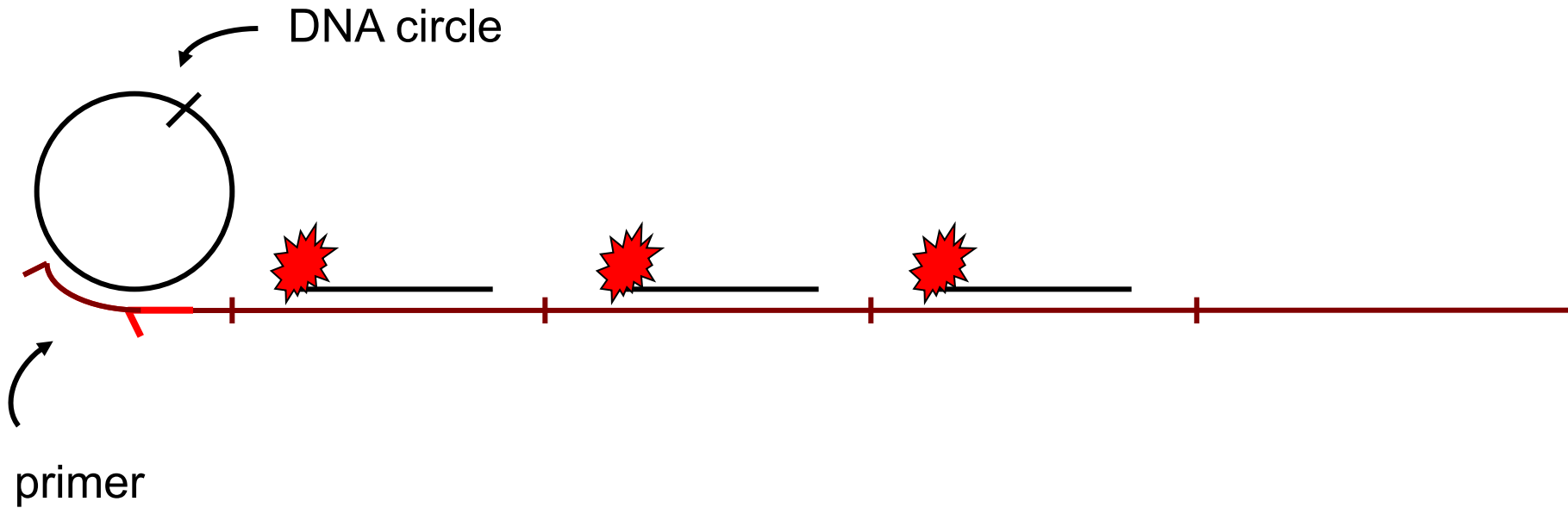


C Wobble

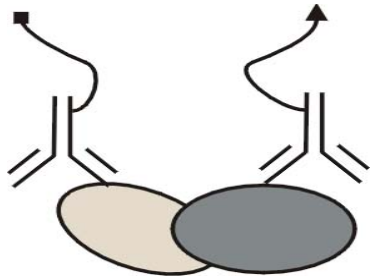




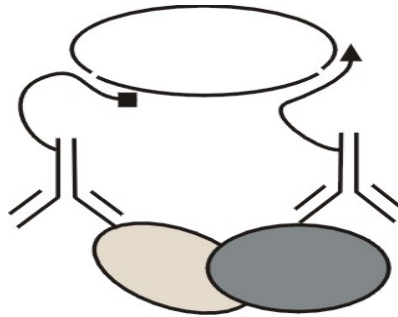
Rolling Circle Amplification



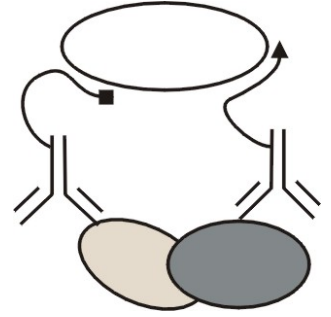
in situ Proximity Ligation Assay (*in situ* PLA)



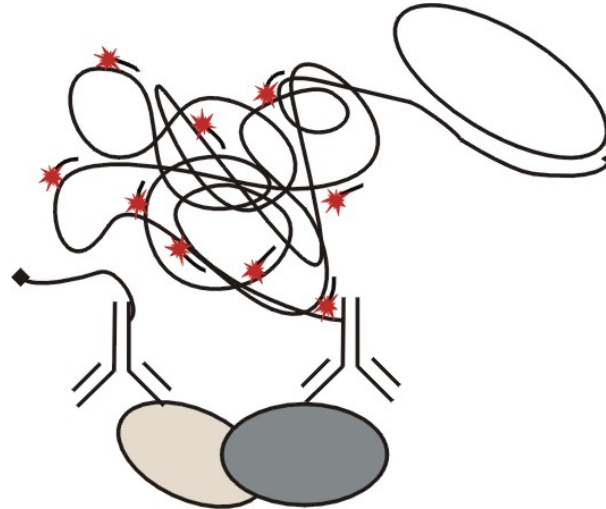
Proximity probes



Circularization probes



Ligation



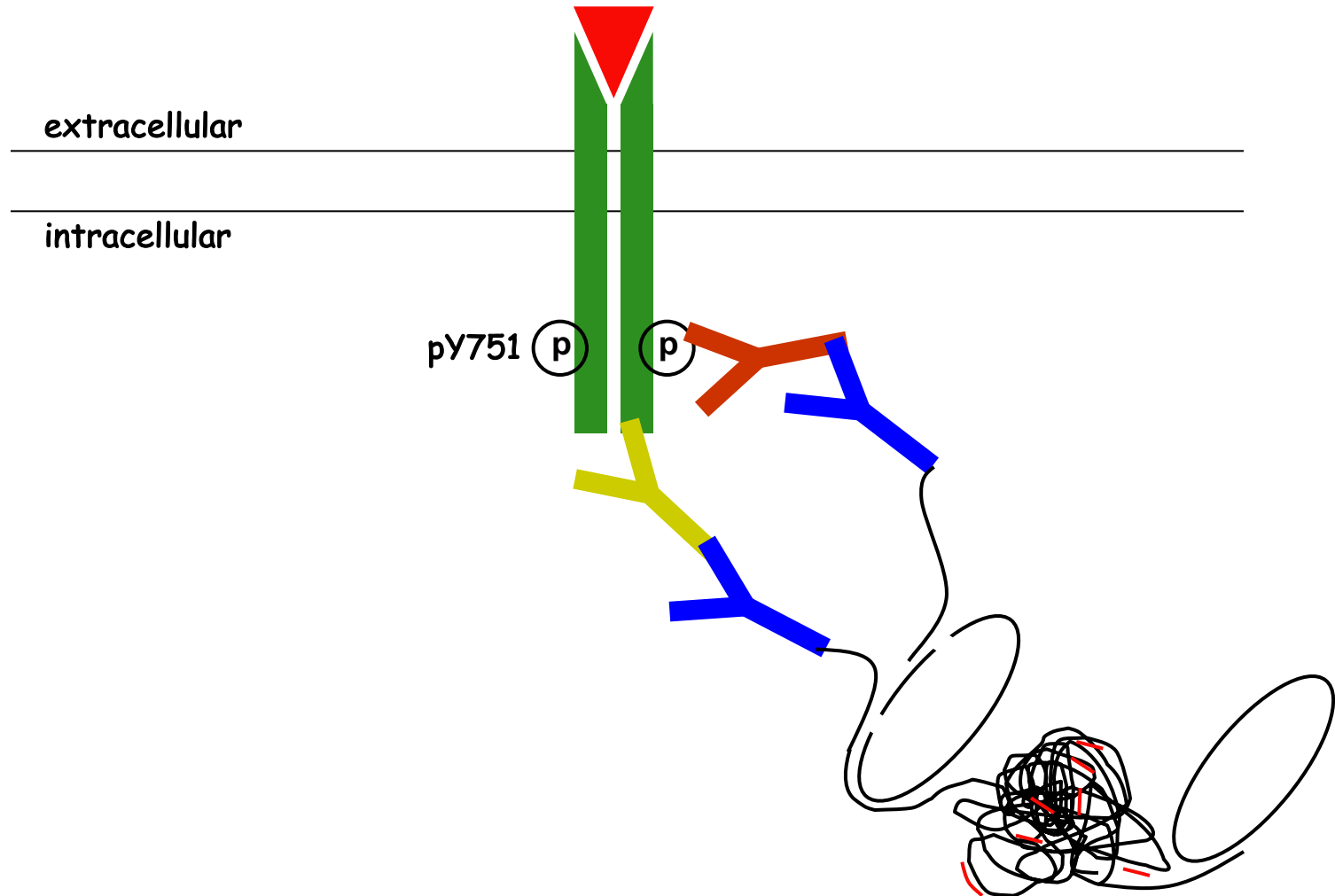
RCA & Detection



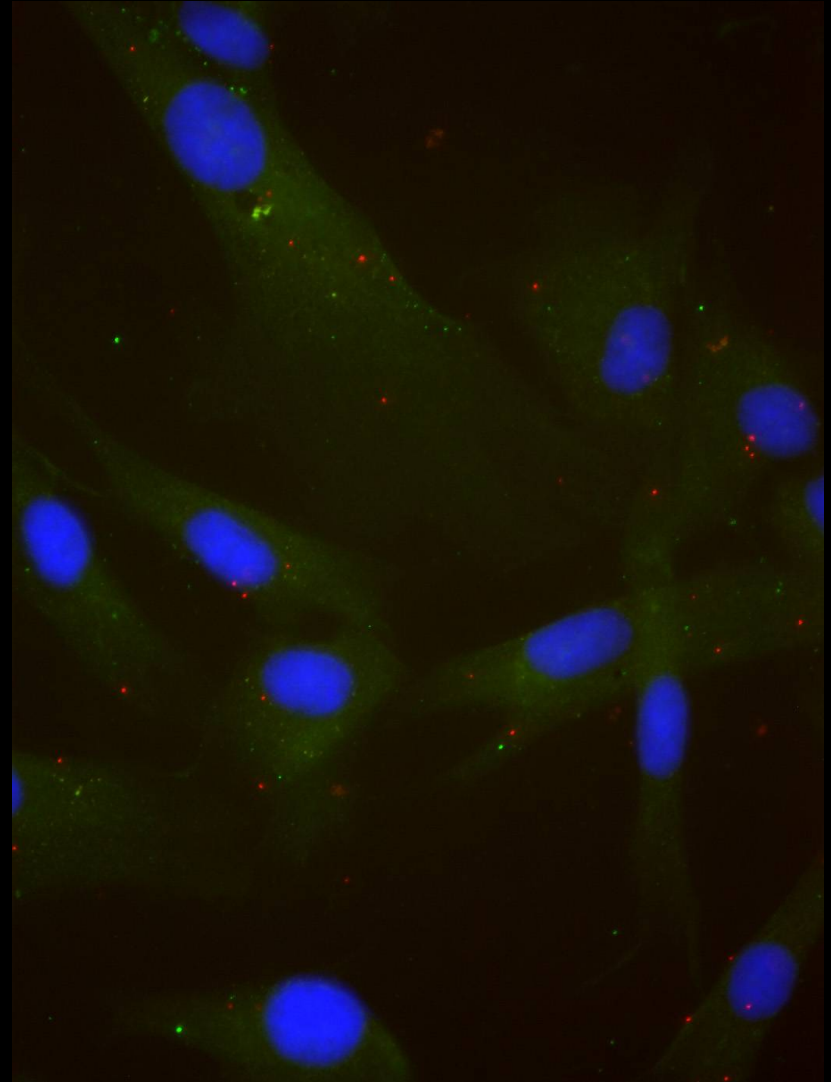
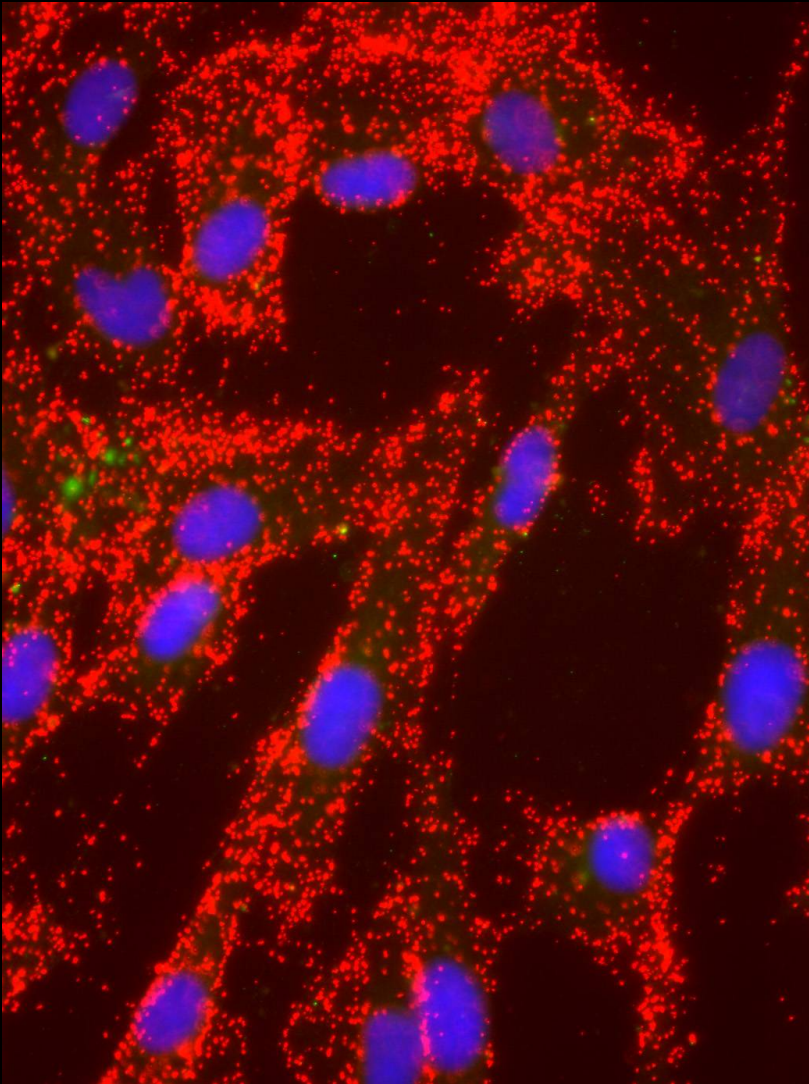
CMYC/MAX interactions



Detection of phosphorylated PDGFR β (secondary proximity probes)

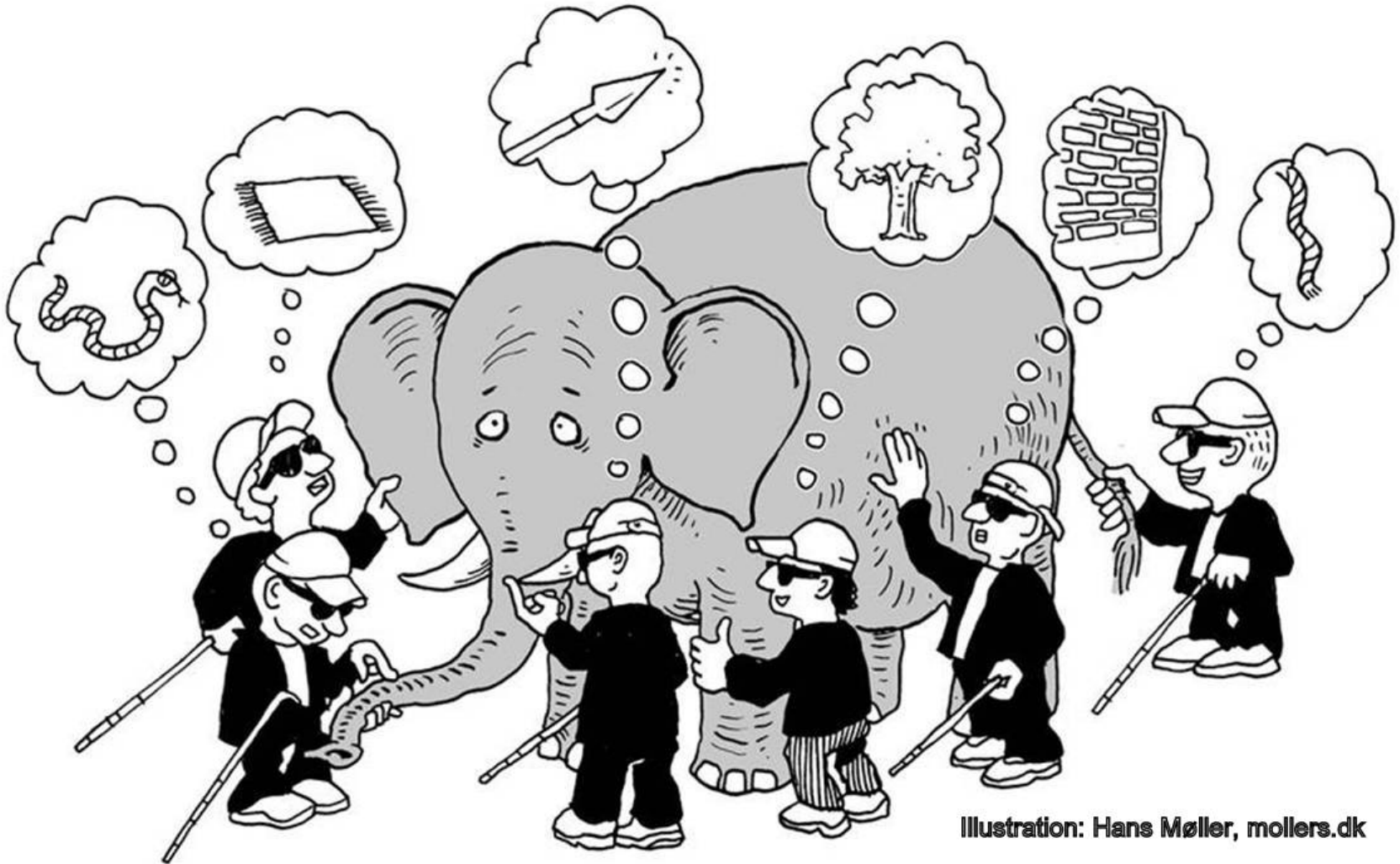


Detection of phosphorylated PDGFR β in human Fibroblasts by *in situ* PLA

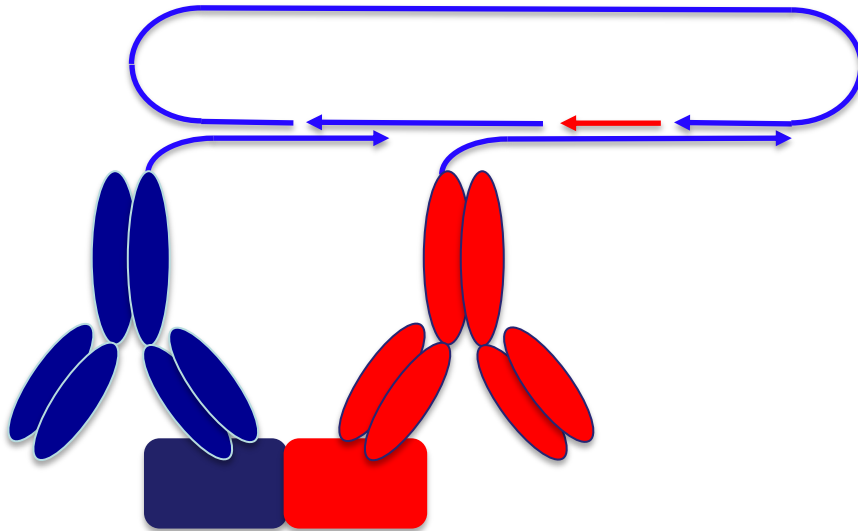
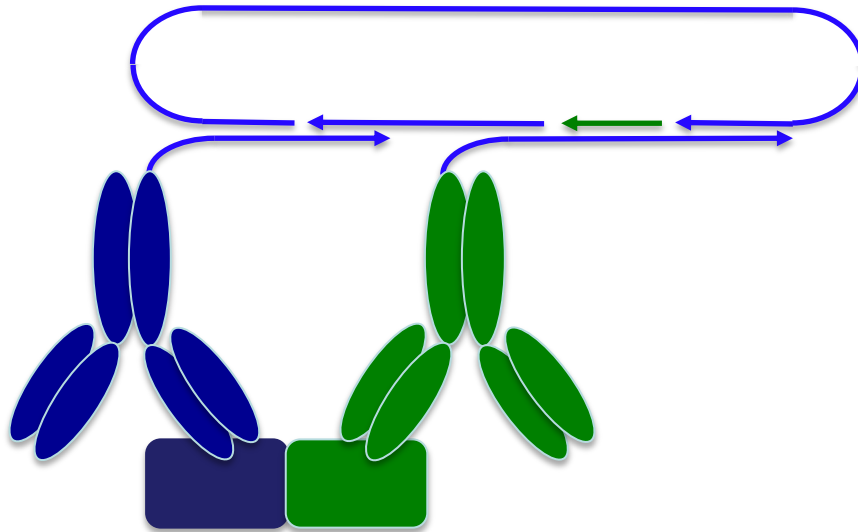


Räcker det med att detektera en
protein interaktion för att förstå
biologin?

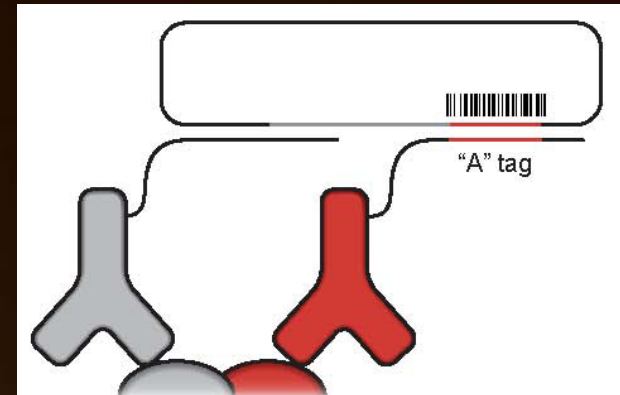
Multiplexing



Multiplex *in situ* PLA



Multiplex *in situ* PLA

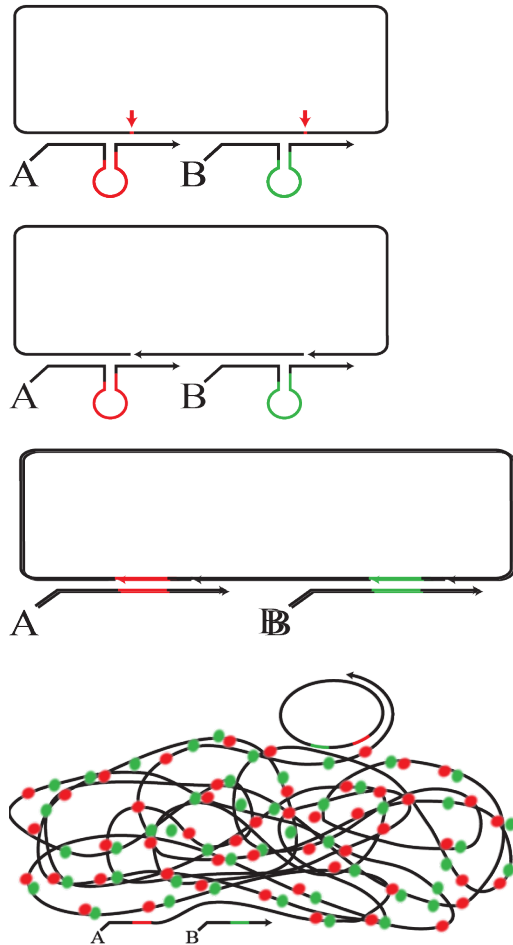


Her2-EGFR
Her2-Her2
Her2-Her3

Molecular Boolean analysis - MolBoolean

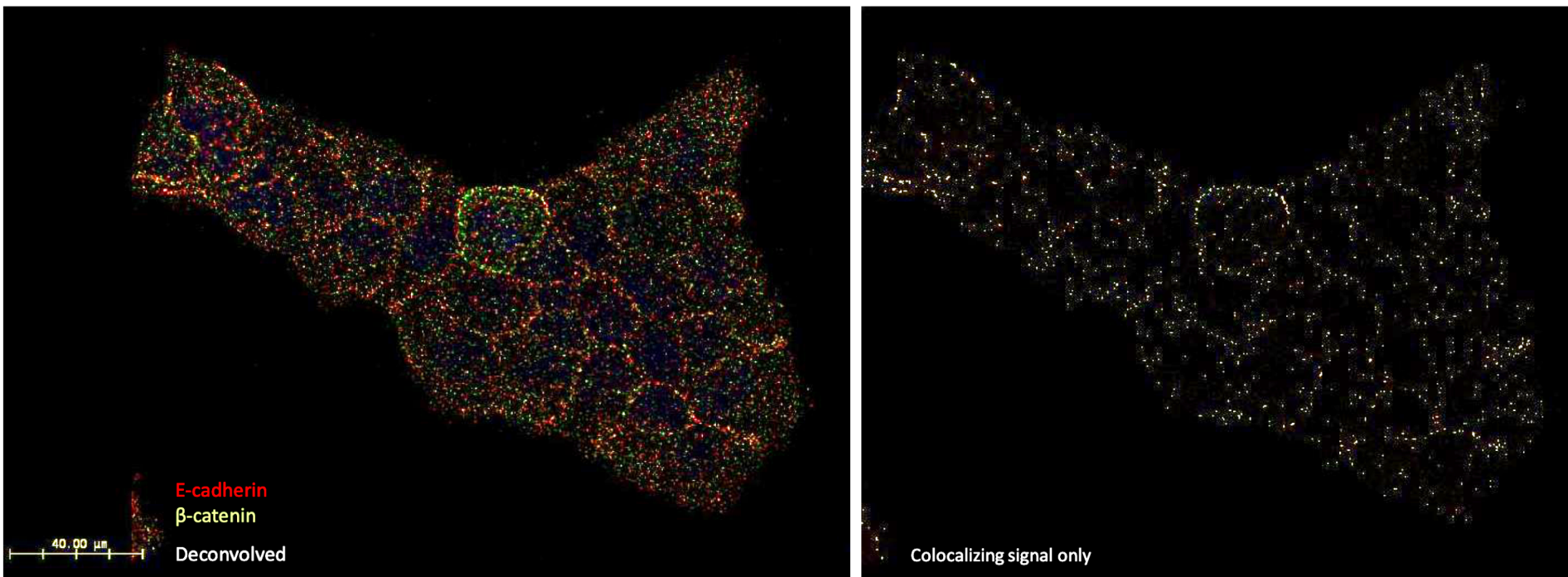
$$KD = \frac{[A][B]}{[AB]}$$

Molboolean design

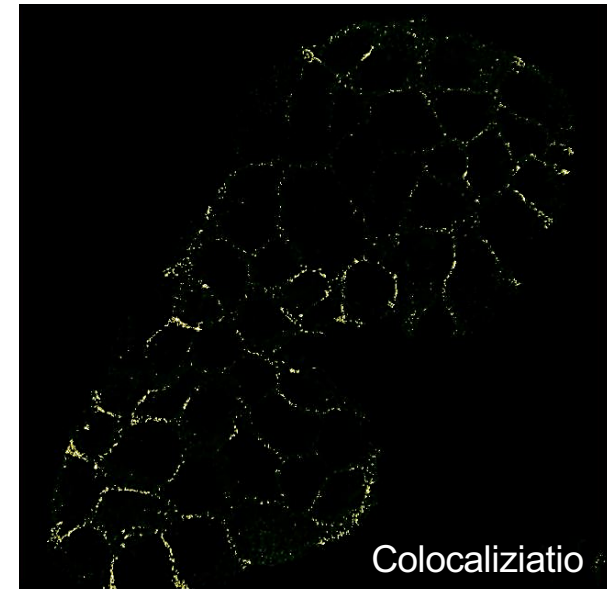
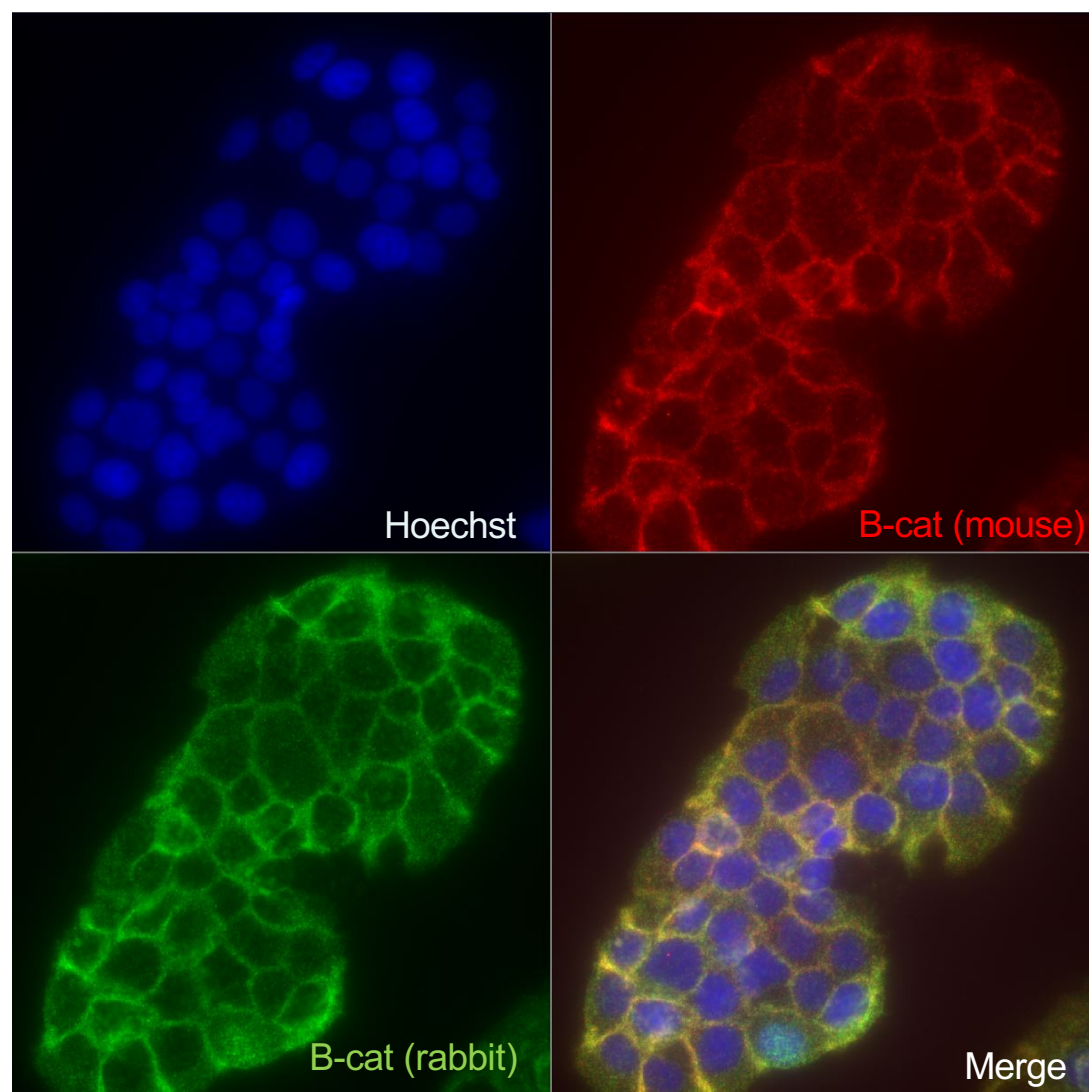


- ➔ • Probe binding
- ➔ • Circle hybridization
- ➔ • Digestion with Nickase
- ➔ • Tag hybridization and ligation
- ➔ • Rolling circle amplification
- Detection

Molboolean analysis of E-Cadherin / β -Catenin



Pairs of antibodies targeting different epitopes



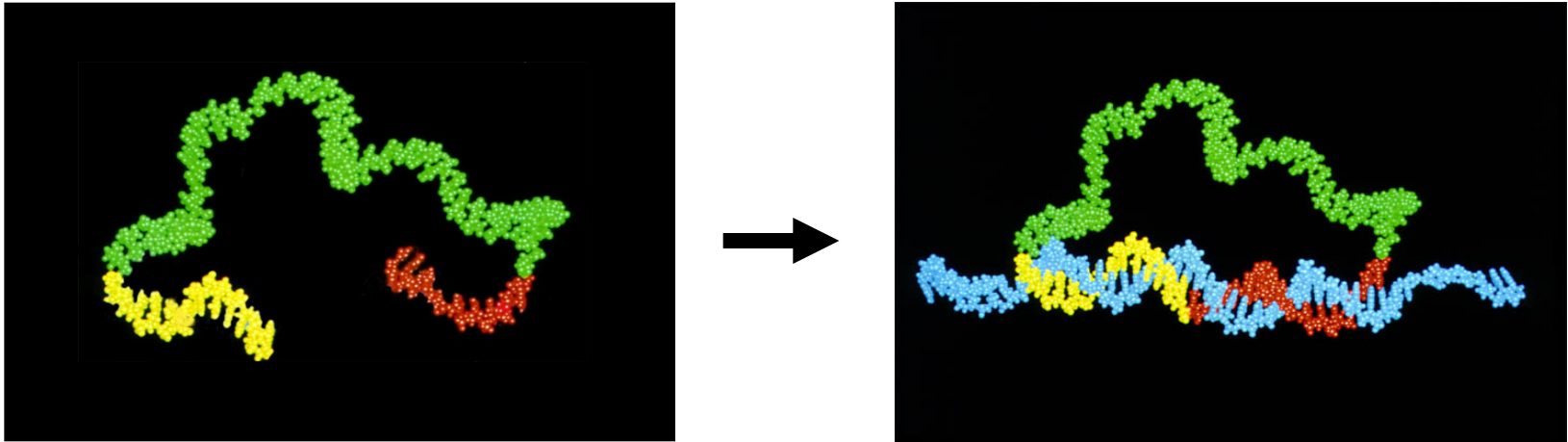
Pearson: 0,604

Raykova & Kerpatsou *et al.*,
unpublished

Räcker det med att detektera
protein interaktioner för att förstå
biologin?

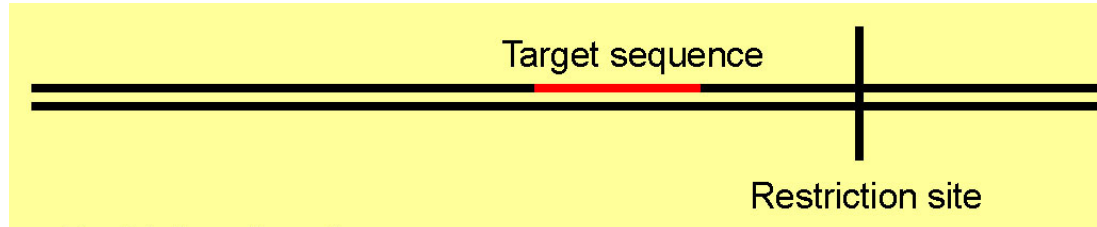
Kan vi kombinera olika slags
metoder för att få en djupare
förståelse?

Padlock probes

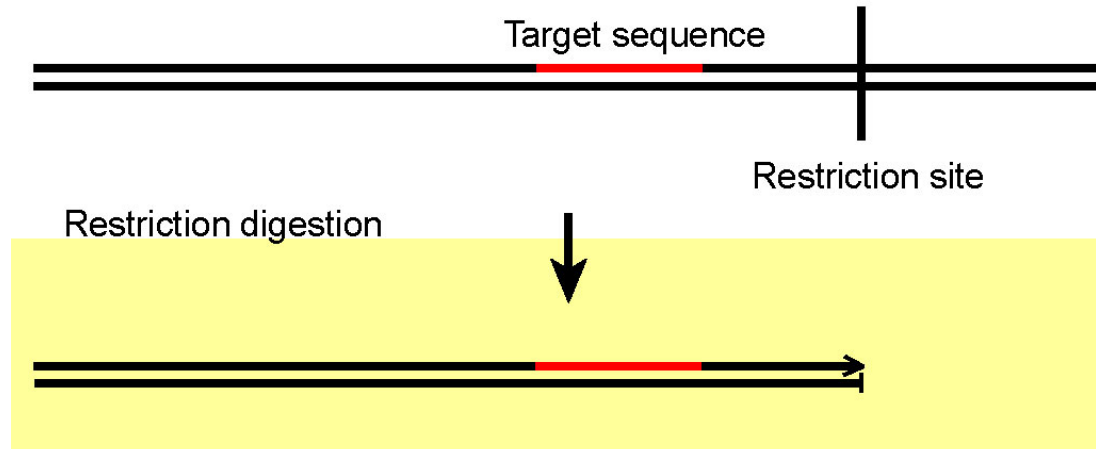


Nilsson *et al.*, *Science* **265**:2085 (1994).
Nilsson *et al.*, *Nature Genetics* **16**:252 (1997).
Nilsson *et al.*, *Nature Biotechnol* **18**:791 (2000).

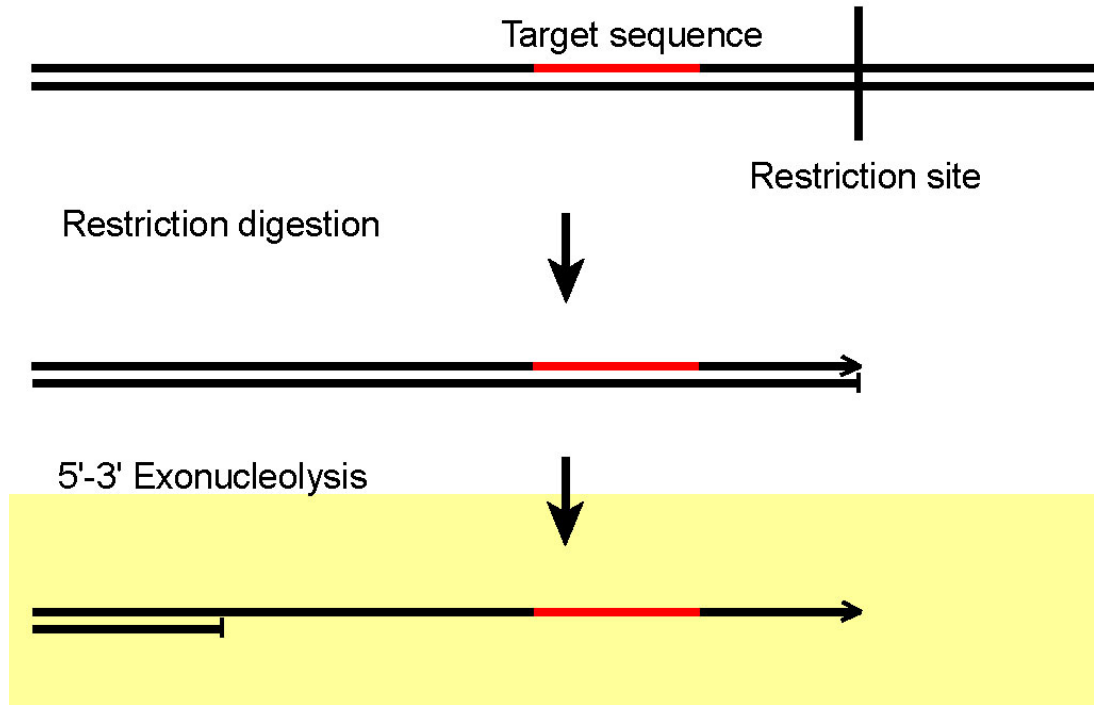
Target preparation – step 1



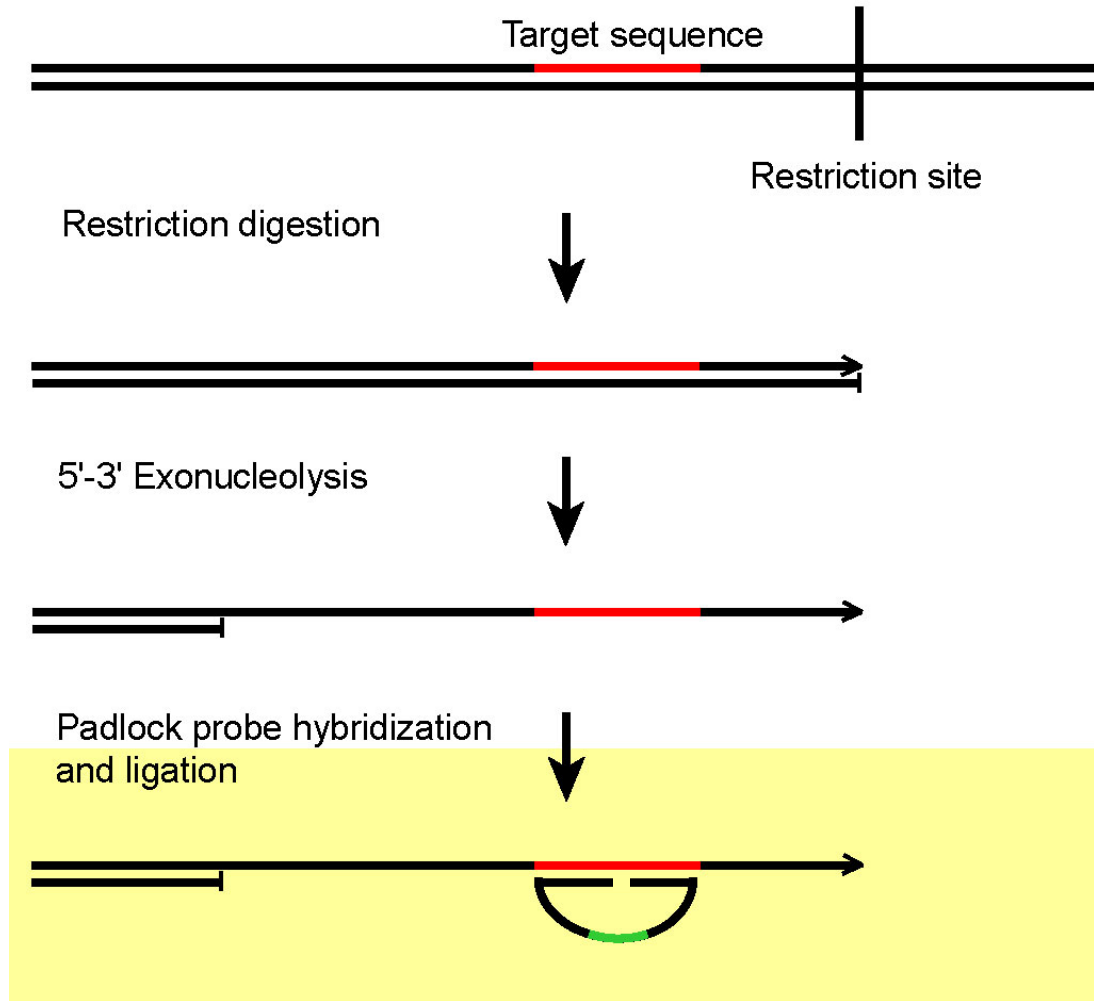
Target preparation – step 2



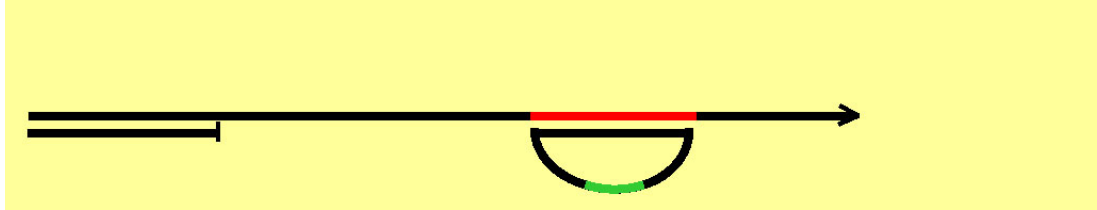
Target preparation – step 3



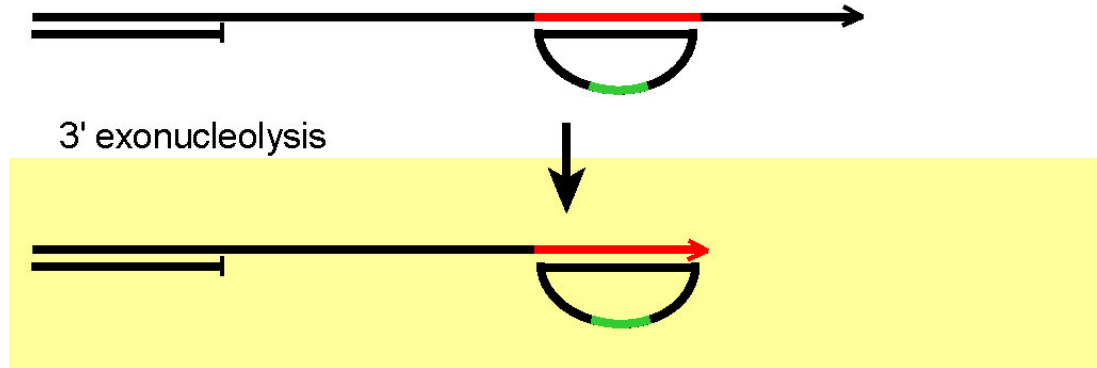
Target recognition by padlock probe



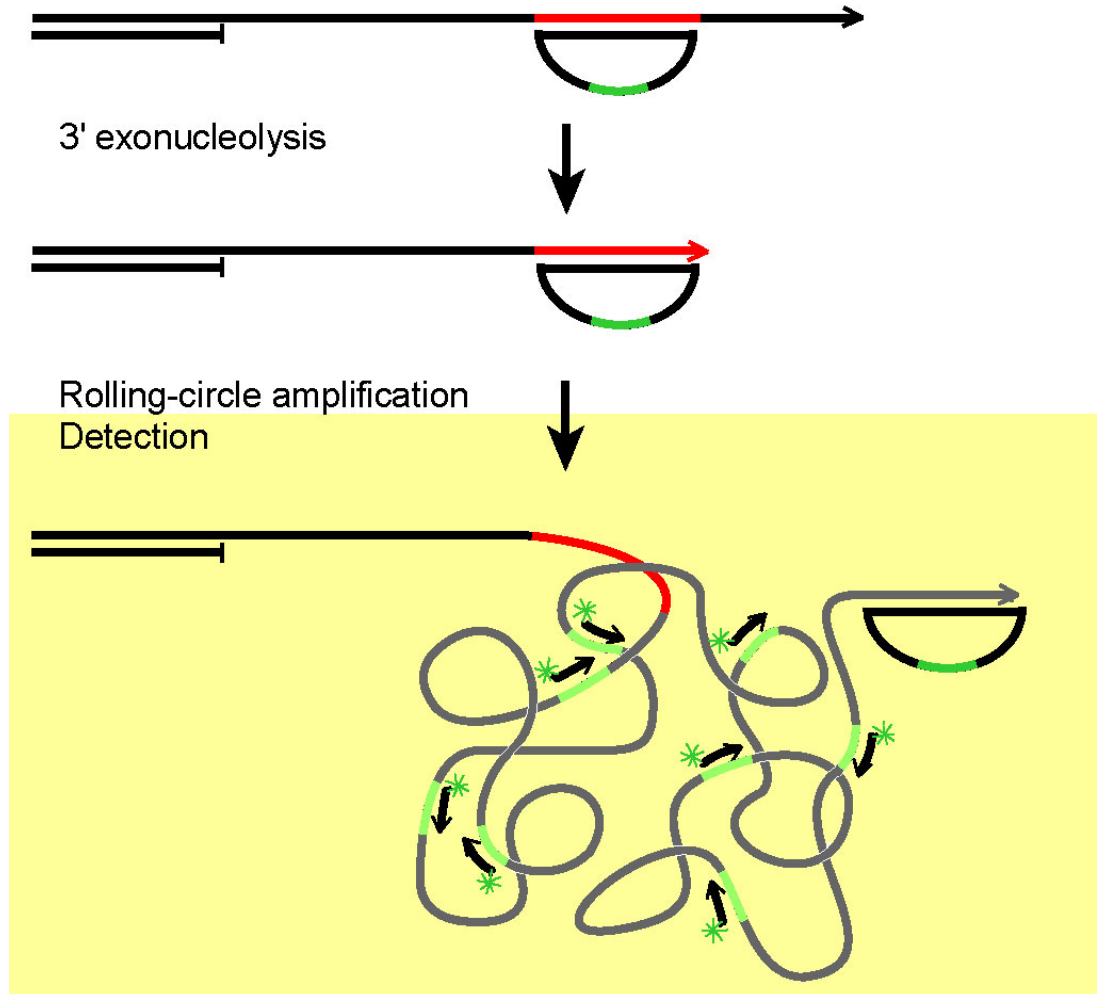
Target-primed RCA



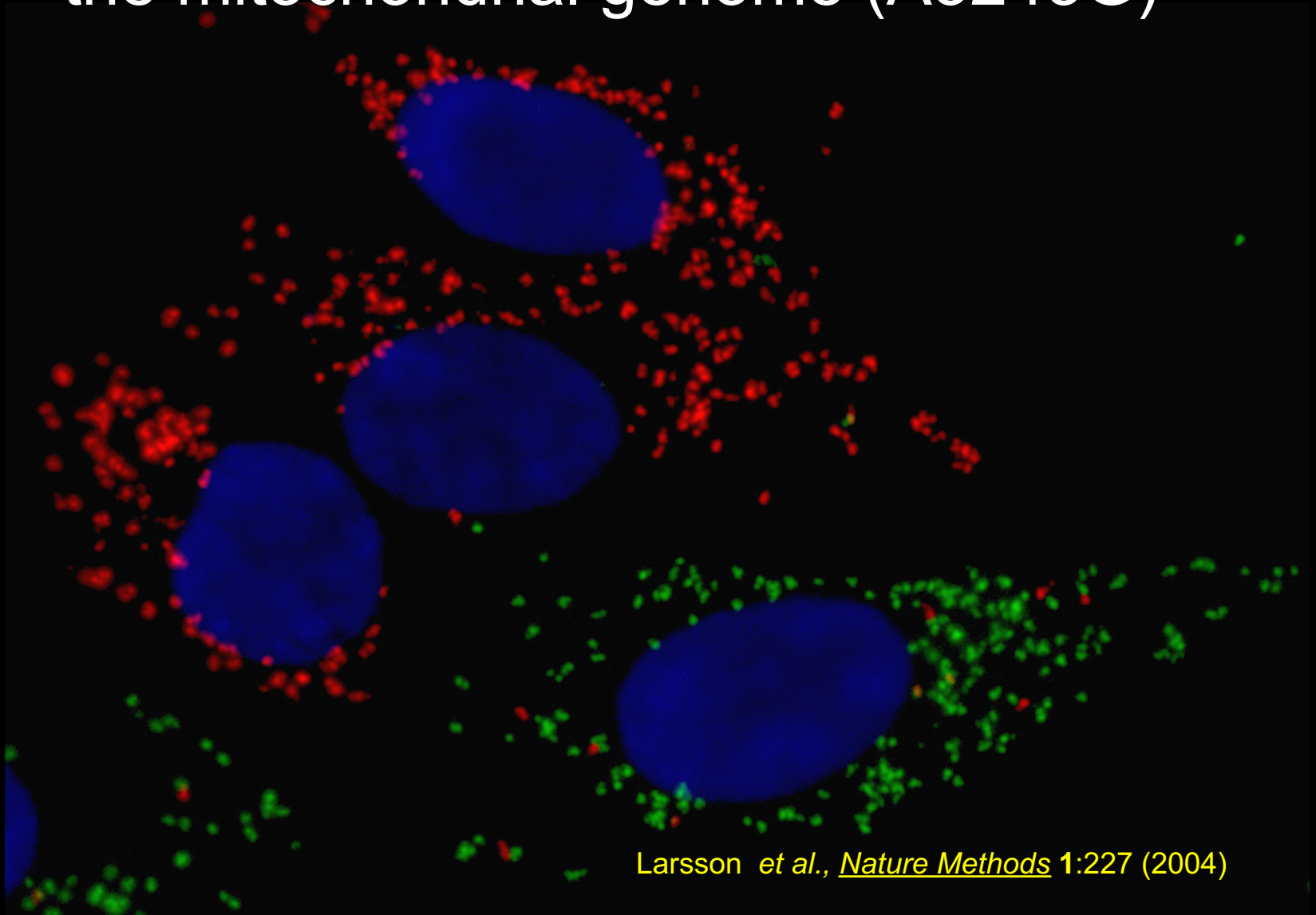
Target-primed RCA



Target-primed RCA

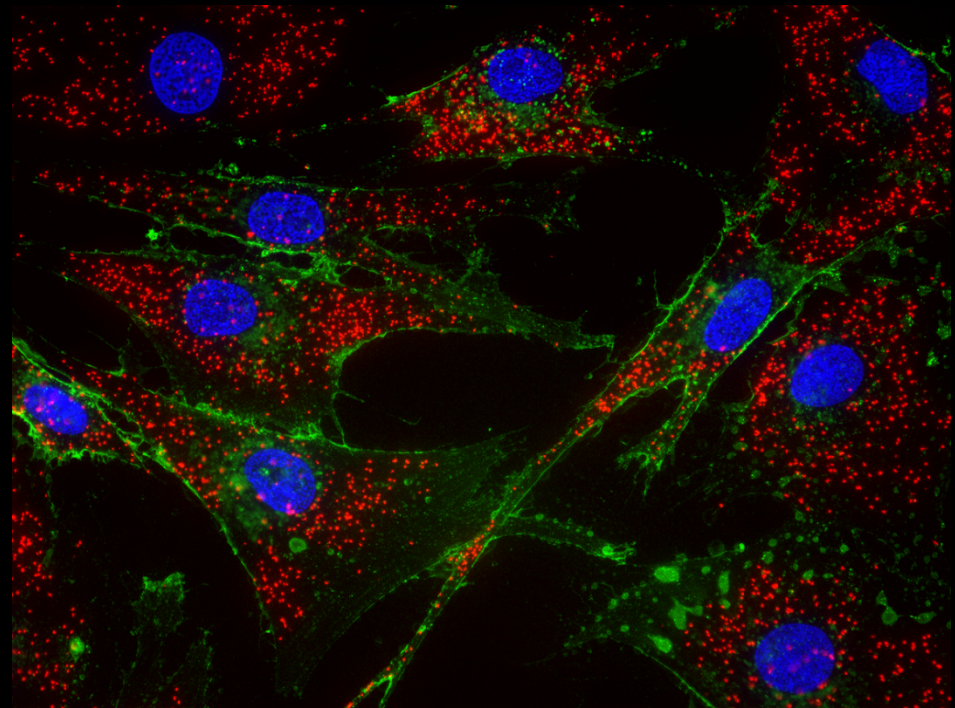
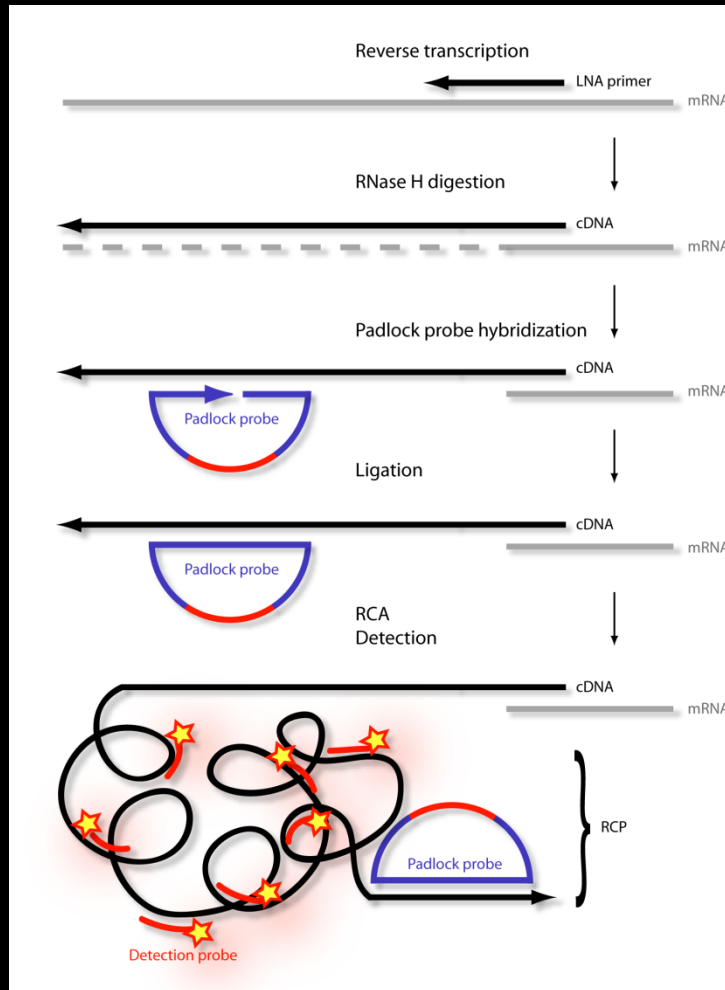


Detection of single nucleotide variation in the mitochondrial genome (A3243G)



Larsson et al., *Nature Methods* 1:227 (2004)

Detection of single transcripts using padlock probes and RCA



β -actin transcripts
Nuclear staining
Membrane staining

30% detection efficiency

Larsson, C. *et al.* (2010) *Nature Methods*, 7, 395–397.

K-RAS genotyping of mRNA

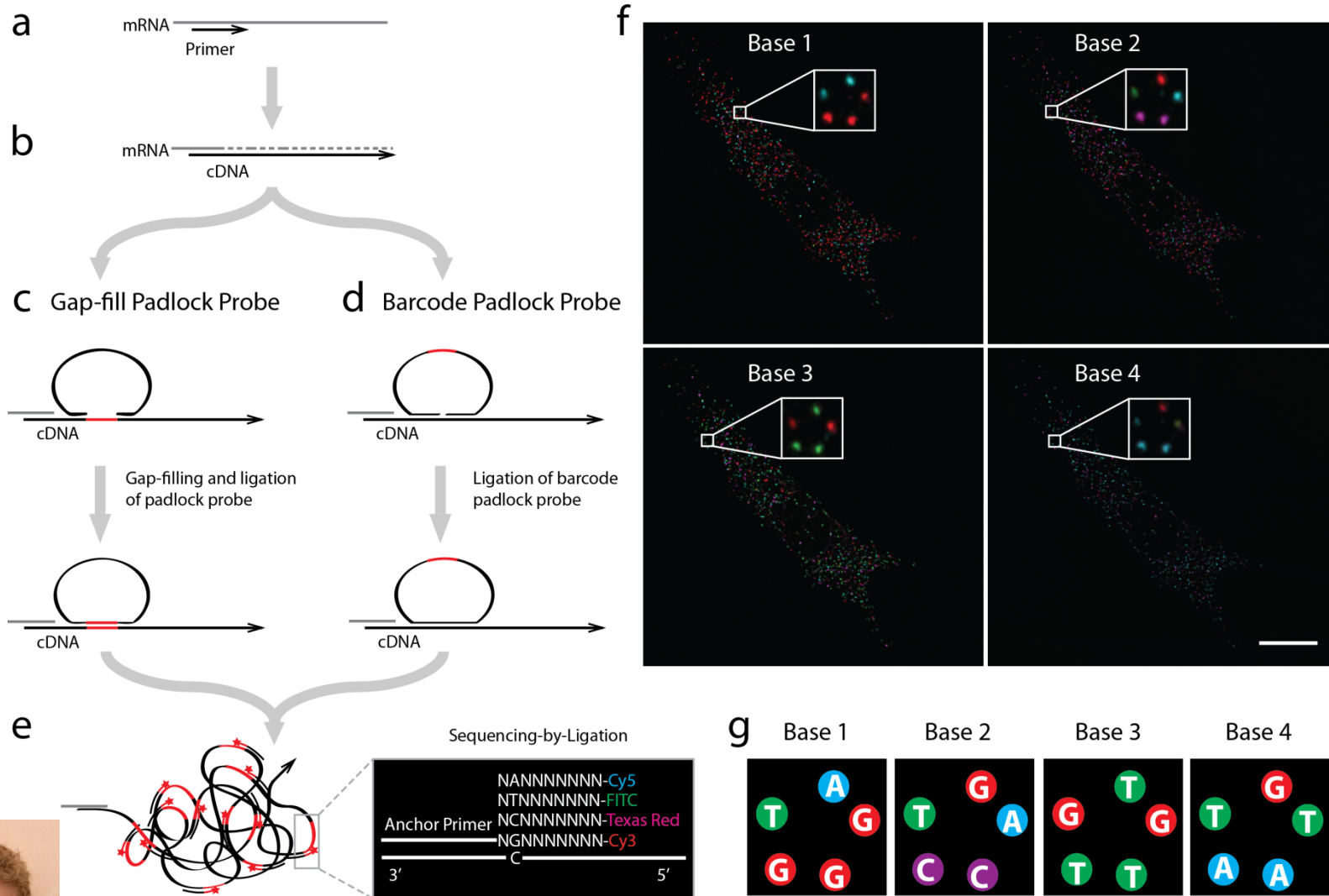
Homozygots wt GGT

Homozygots GTT

Heterozygots GGT, GAT

Co-culture homozygots
GGT, GTT

Schematic Illustration of *in situ* Sequencing

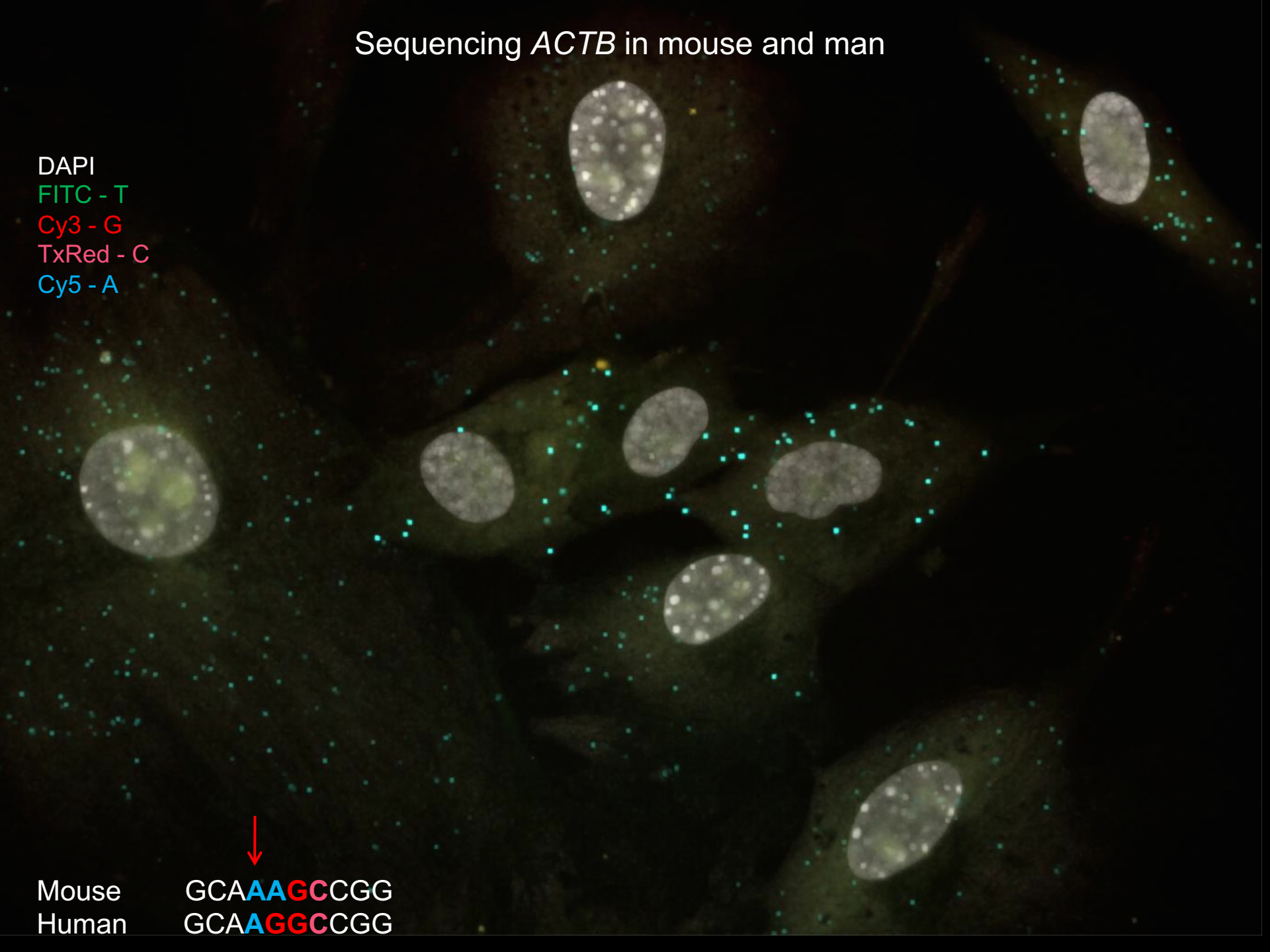


Ke, et al, *Nature Methods*, 10:857 (2013)

Sequencing *ACTB* in mouse and man

DAPI
FITC - T
Cy3 - G
TxRed - C
Cy5 - A

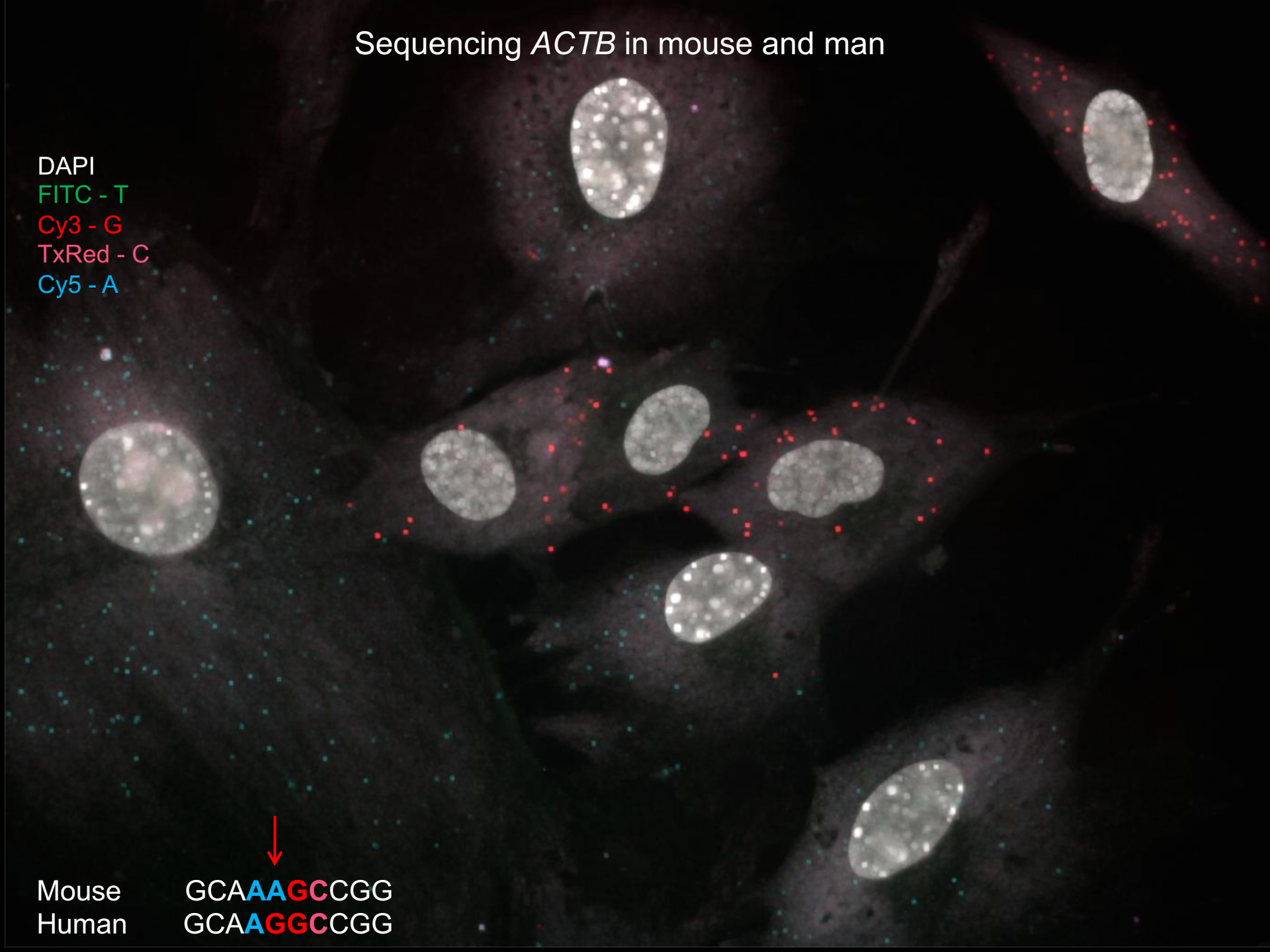
↓
Mouse GCA**A****G**CCGG
Human GCA**A****G**CCGG



Sequencing *ACTB* in mouse and man

DAPI
FITC - T
Cy3 - G
TxRed - C
Cy5 - A

↓
Mouse GCA**A**GC**C**CGG
Human GCA**G**GC**C**CGG



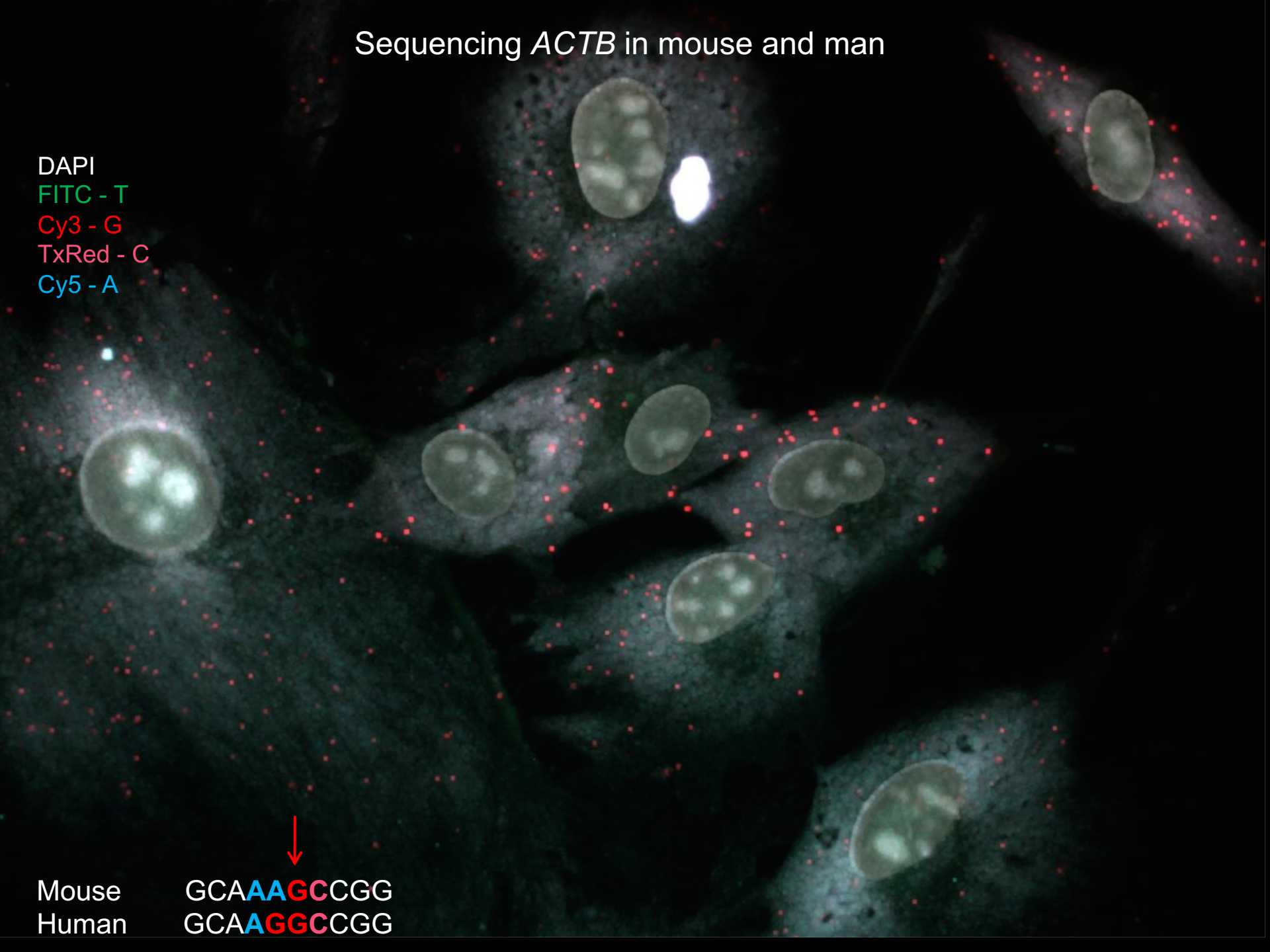
Sequencing *ACTB* in mouse and man

DAPI
FITC - T
Cy3 - G
TxRed - C
Cy5 - A

Mouse
Human

↓

GCA**A**GC**C**CGG
GCA**A**GC**C**CGG



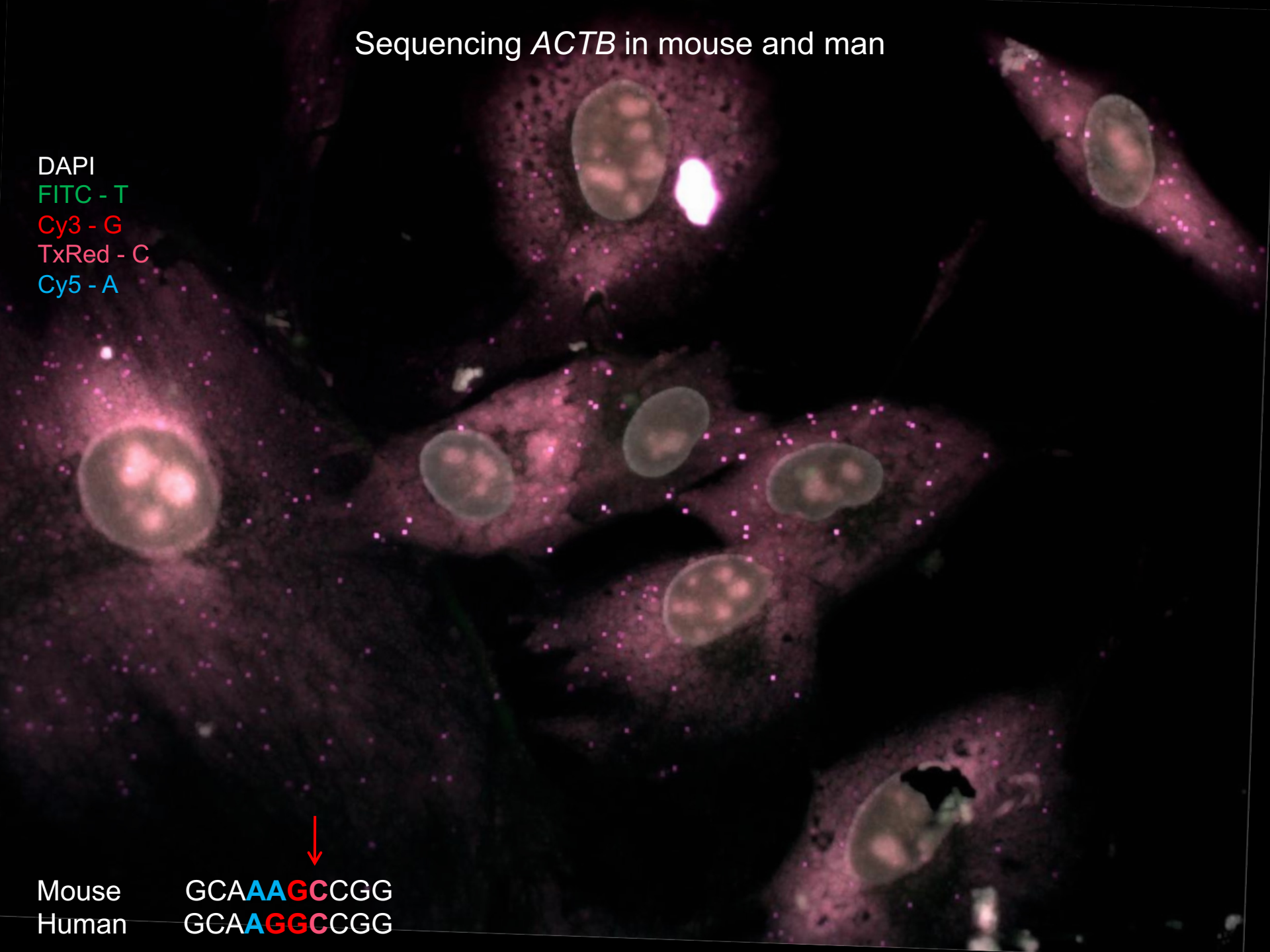
Sequencing *ACTB* in mouse and man

DAPI
FITC - T
Cy3 - G
TxRed - C
Cy5 - A

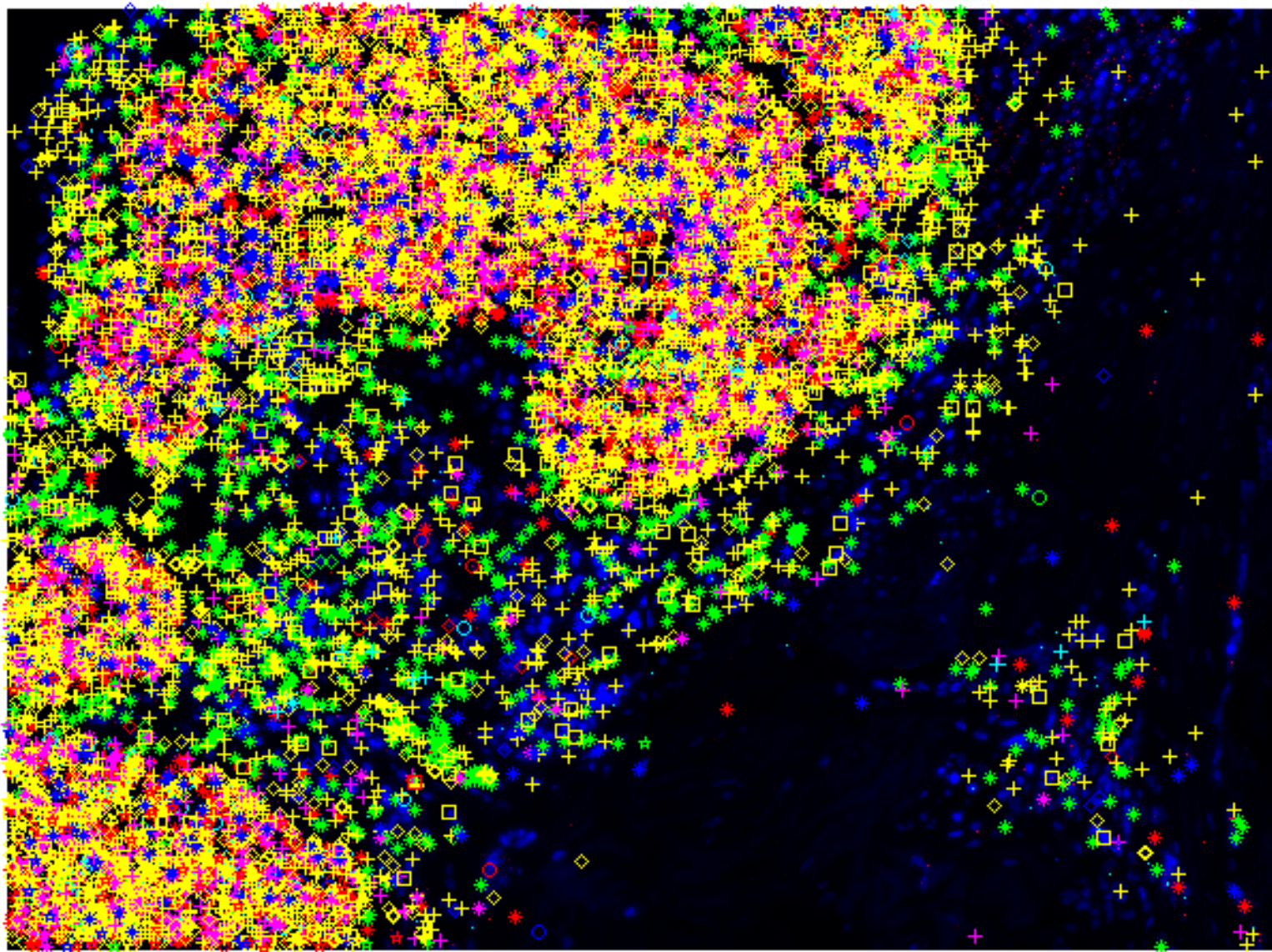
Mouse
Human

GCA**A**AGCCGG
GCA**A**GGCCGG

↓

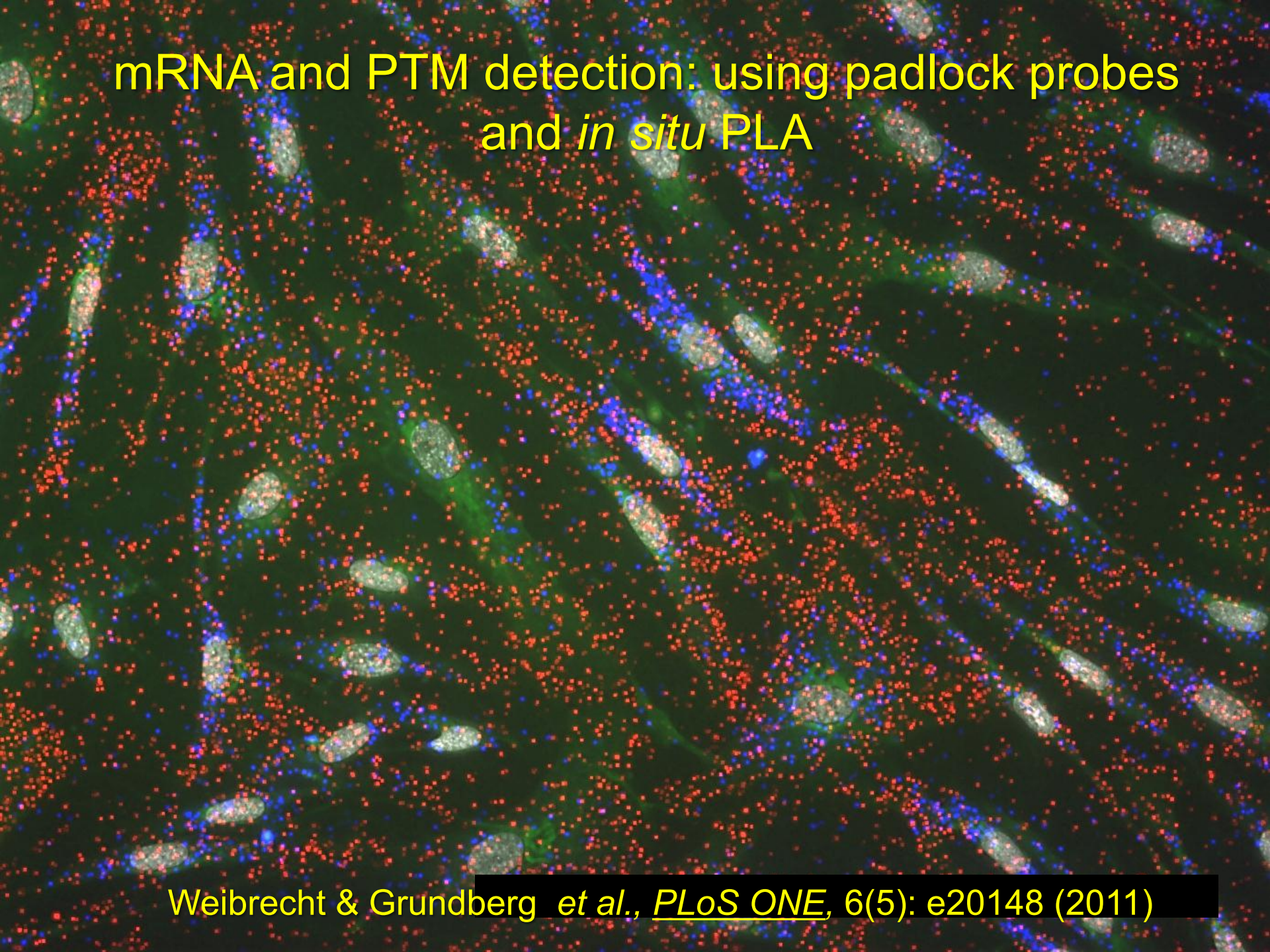


Expression profiling



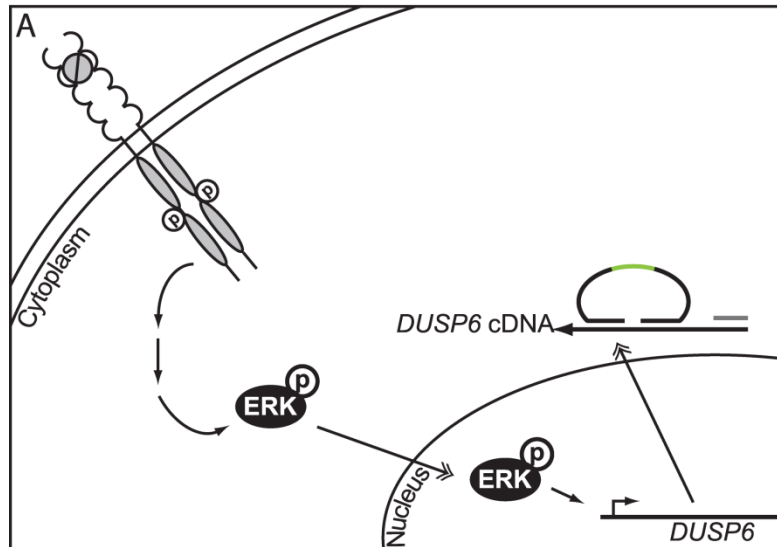
- * HER2
- * VIM
- KI-67
- STK 15
- BIRC5
- CCNB1
- MYBL2
- + ST-3
- * CTSL2
- ◇ ER
- ◇ BAG1
- ◇ CD68
- ◇ BCL2
- + SCUBE2
- ◇ ACTB
- * GAPDH
- * RPLP0
- * GUS
- + TFRC
- + VEGF
- MYC
- MET
- TP53
- EPCAM
- CCND1
- MUC1
- RAC1
- ★ HOXB7
- ★ ATB1
- ★ ETV4

mRNA and PTM detection: using padlock probes and *in situ* PLA

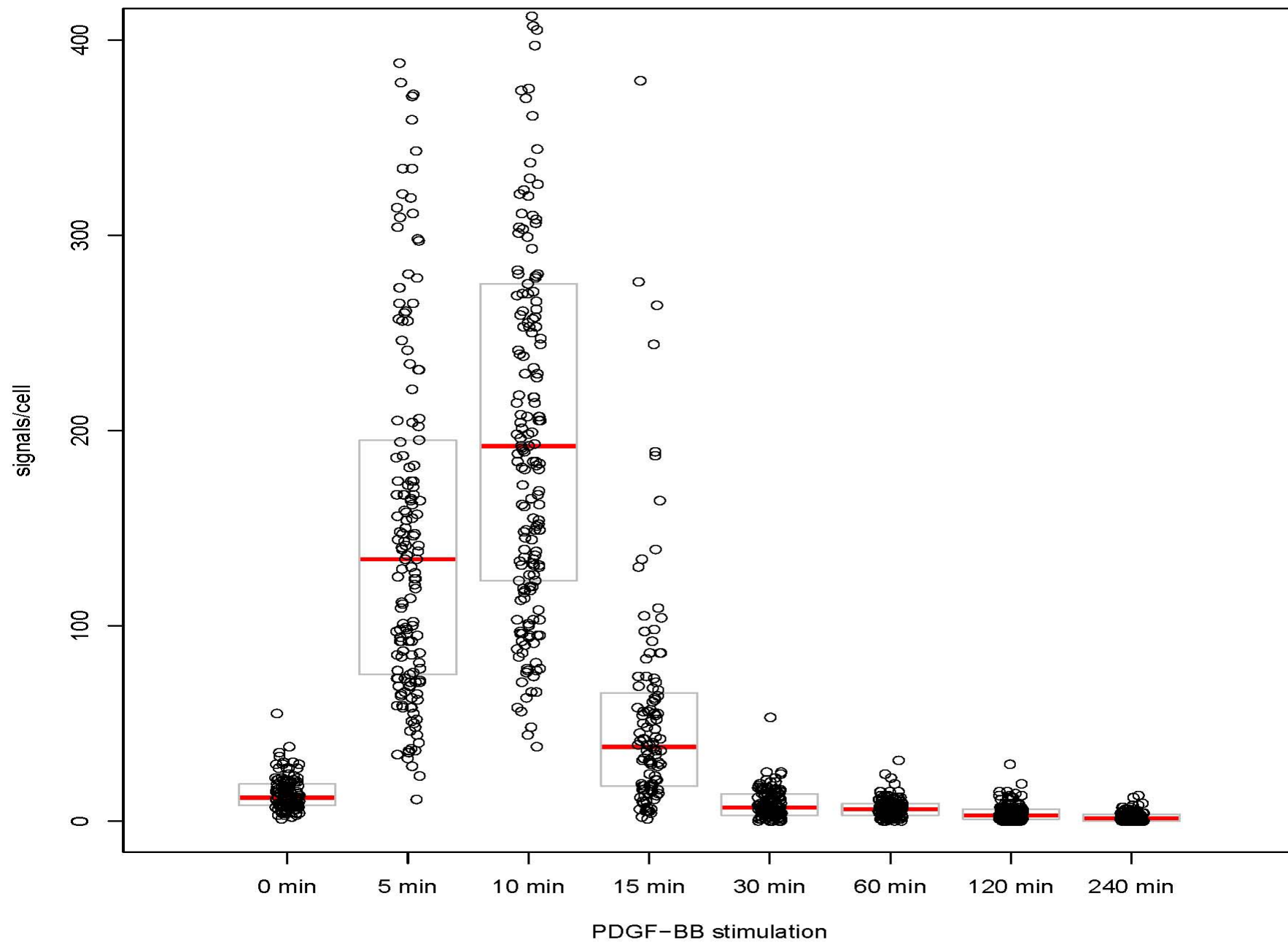


Weibrecht & Grundberg *et al.*, PLoS ONE, 6(5): e20148 (2011)

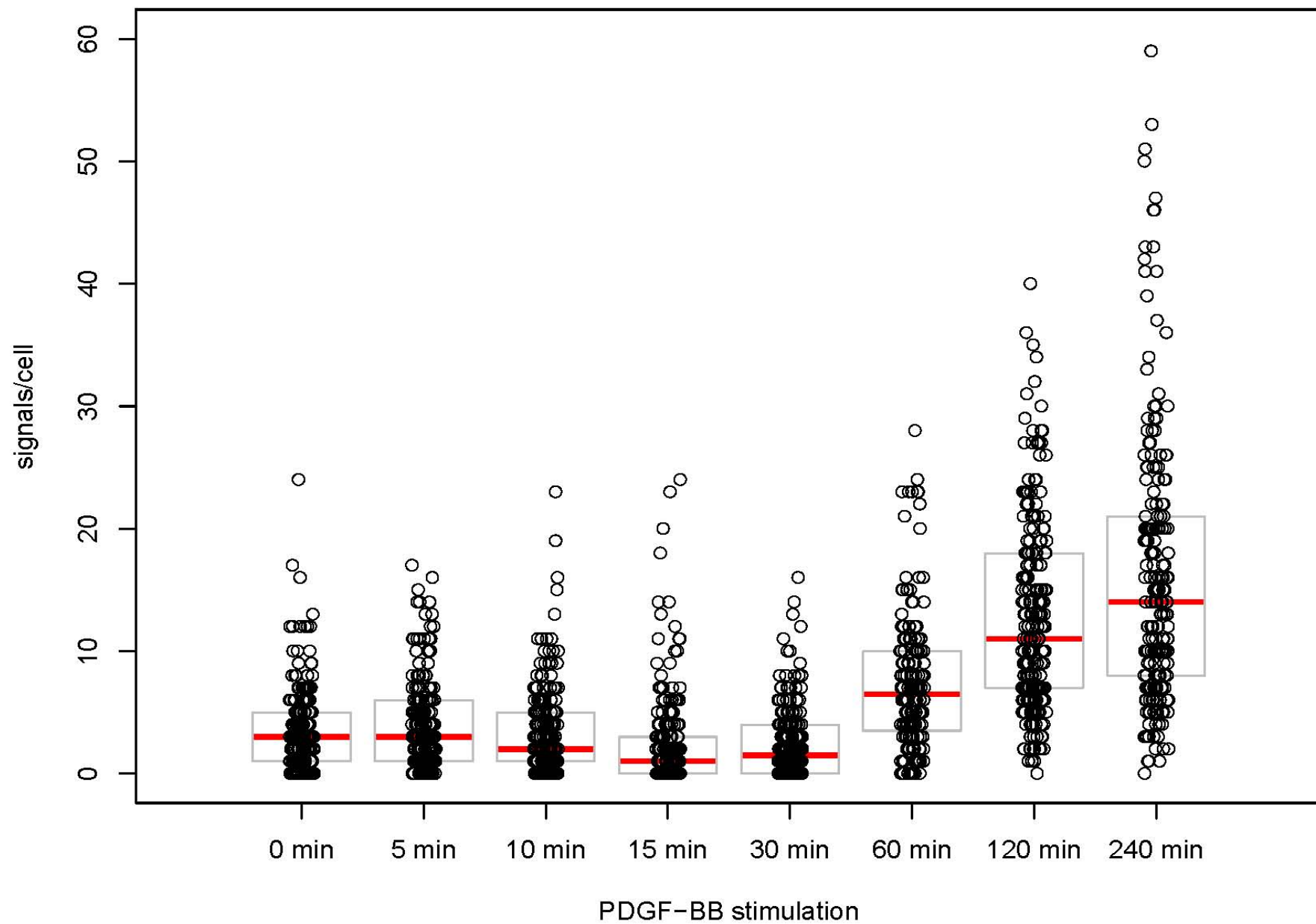
Visualization of signaling activity



in situ PLA for detection of PDGFR-beta phosphorylation



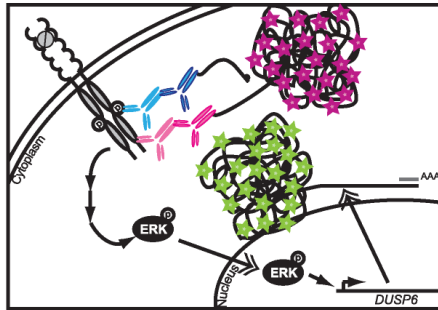
detection of DUSP6 mRNA using padlock probes



Drug-treatment; proof-of-principle

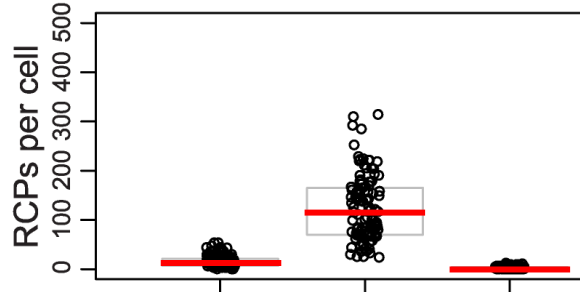
Bi

no drugs

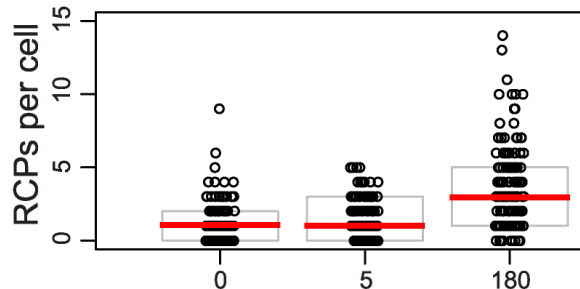


Bii

Simultaneous detection: pPDGFR β



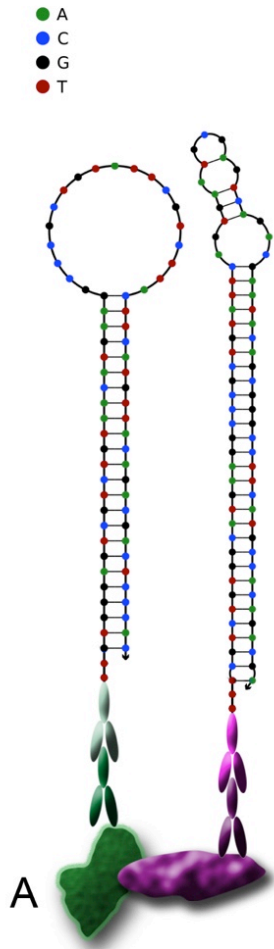
Simultaneous detection: *DUSP6* expression



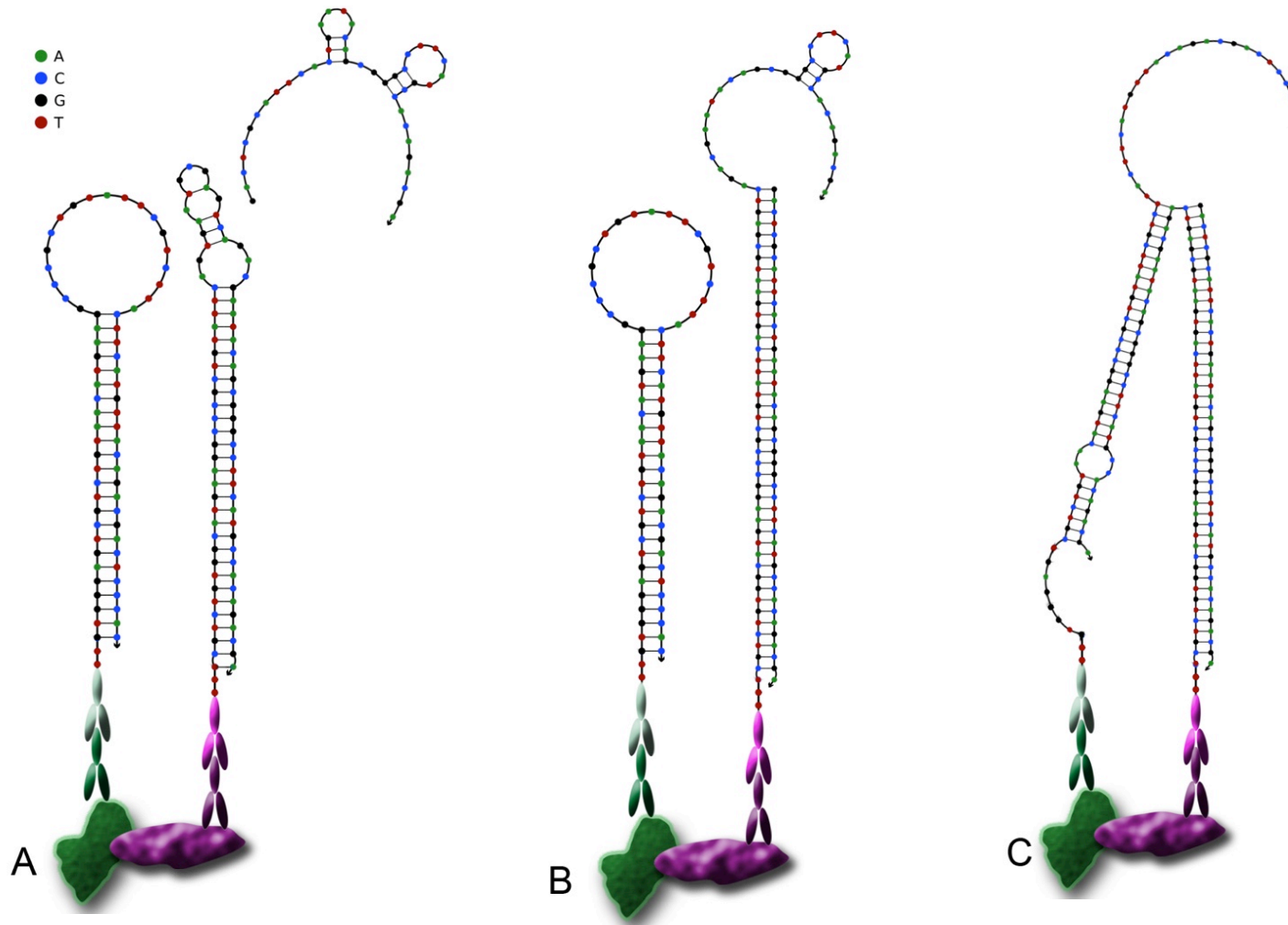
min stimulation with PDGF-BB

Vad kan vi göra billigare och
enklare – så att metoder kan bli
tillgängliga för fler?

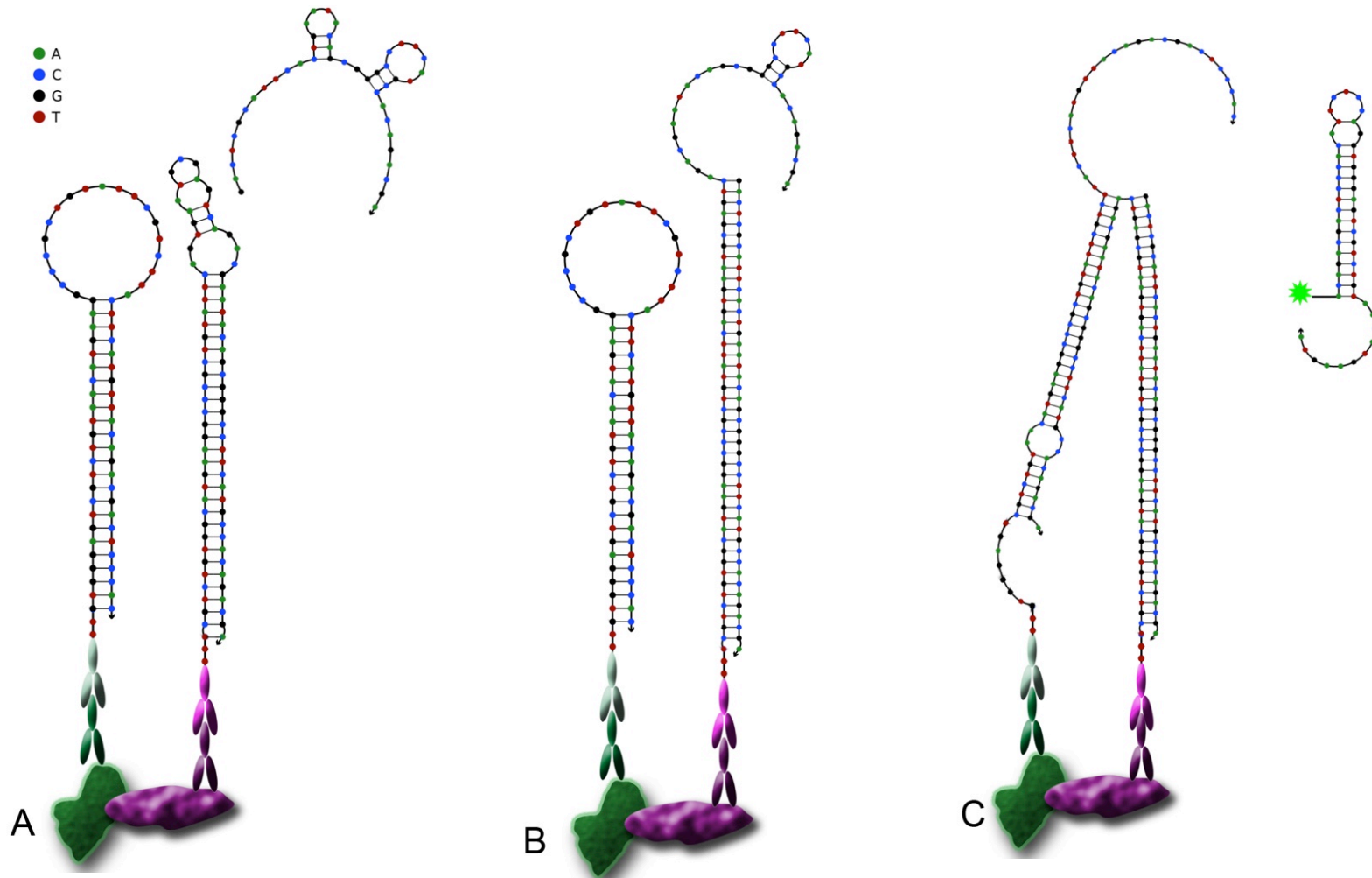
Proximity dependent initiation of HCR



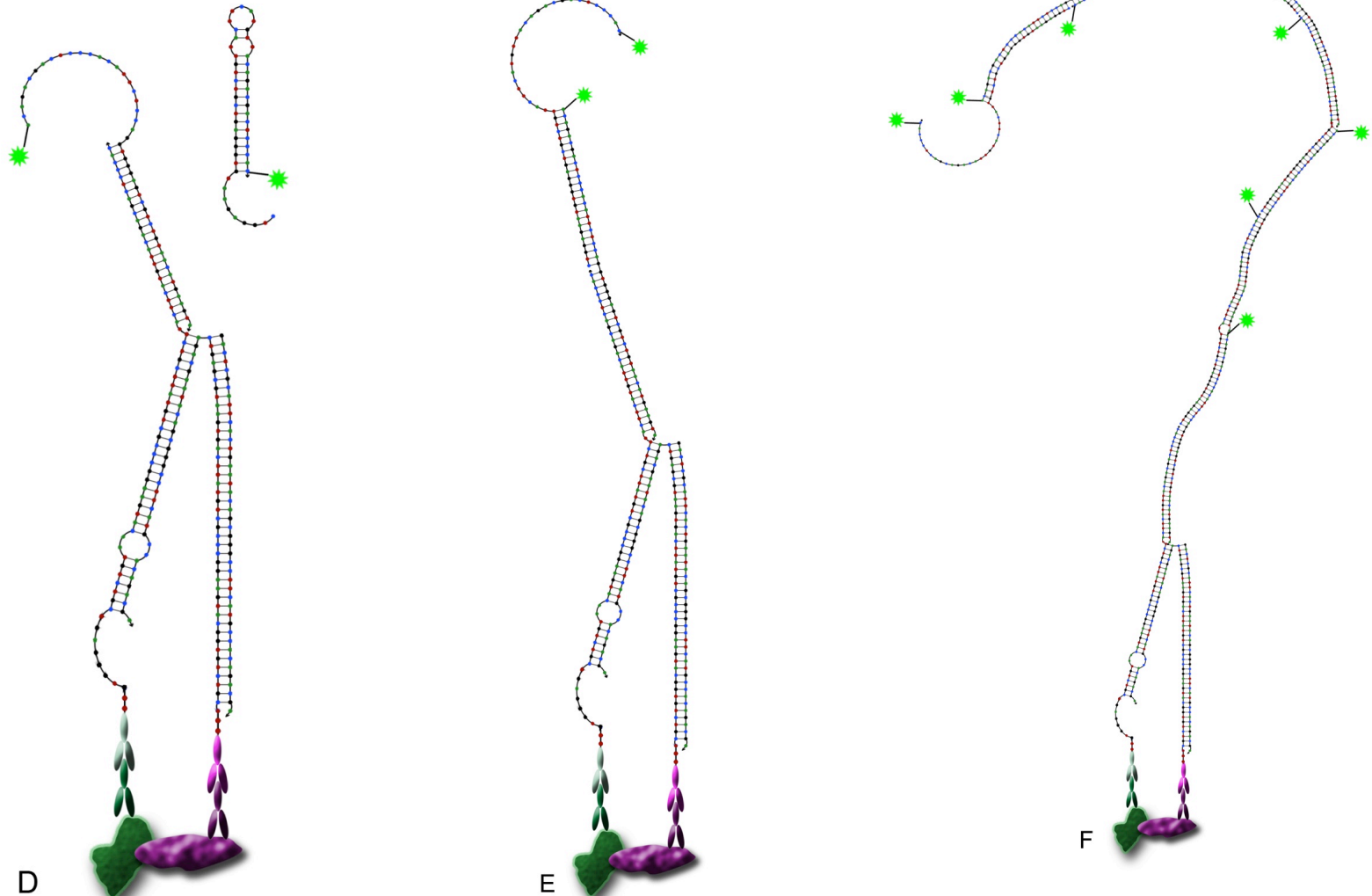
Proximity dependent initiation of HCR



Proximity dependent initiation of HCR

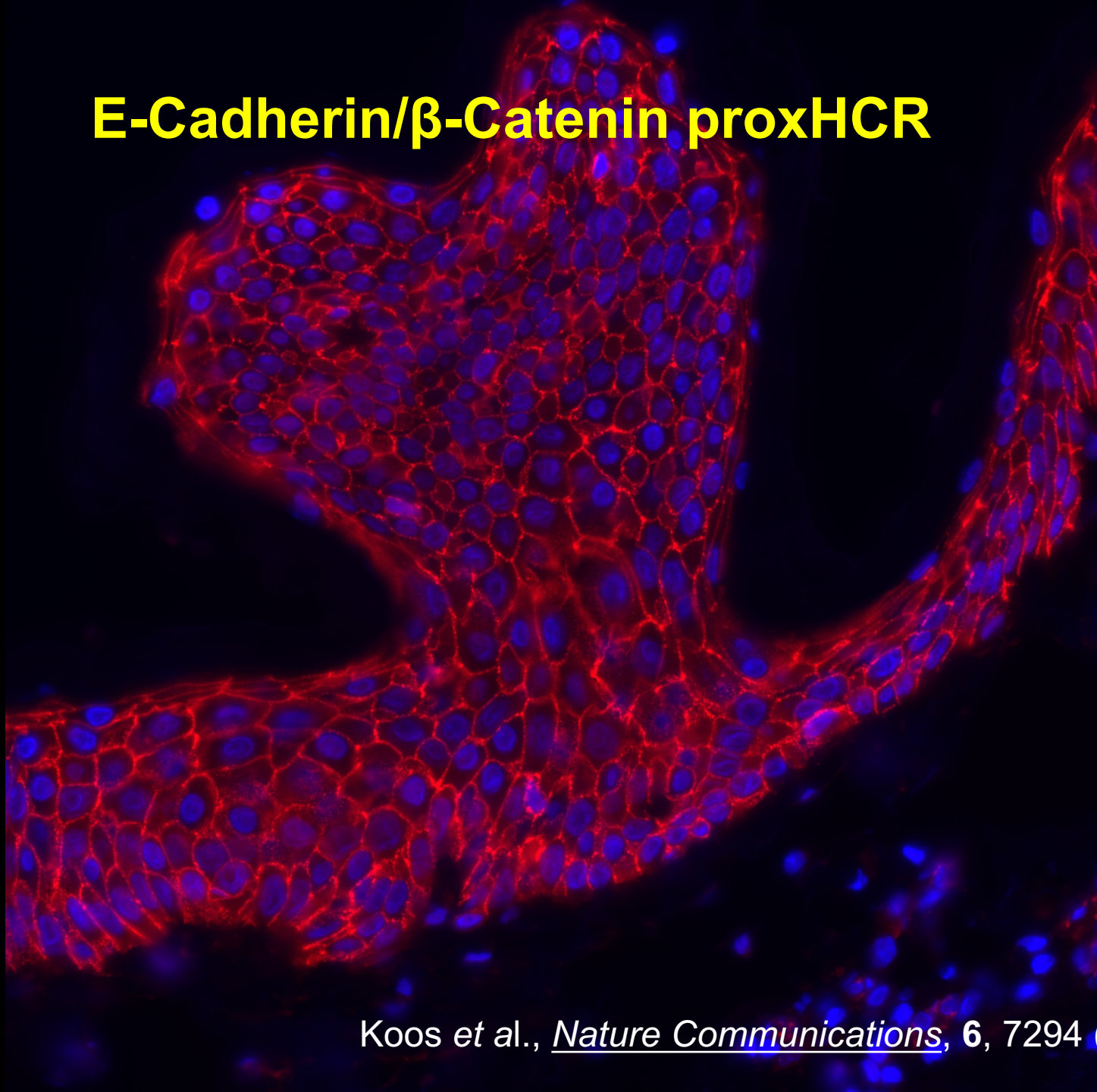


Proximity dependent initiation of HCR



Koos *et al.*, Nature Communications, **6**, 7294 (2015)

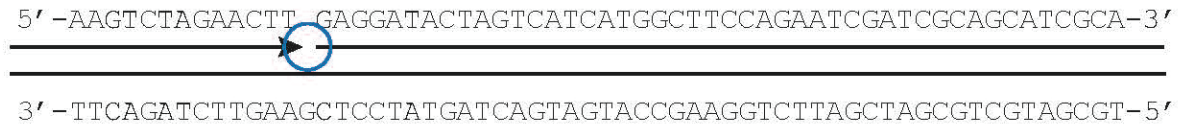
E-Cadherin/ β -Catenin proxHCR



Koos et al., *Nature Communications*, 6, 7294 (2015)

Och nu något helt annorlunda

Identifying DNA nicks



What do you want of a DNA polymerase?

✓ it would need to be highly error prone (possibility to use only 3 nucleotides)

✓ it would

✓ it would
reading)

Sloppymerase™

✓ it would have to have 5' to 3' exonuclease activity to remove the strand in front of the nick while it is synthesizing a new strand

✓ tolerate biotinylated nucleotides

Sloppymerase™ is highly error-prone

Oligo	GGATCCGGCCAAGCTTCGAGCTGAATTCTGCAGTACATTAA
- dCTP	
1. F	GGATAAGGAGAAGGTTTGAGTTGAATTGTGTAGTATATTAA
2. F	GGATTGGGGTGAATTTTGG-----AGTATATTAA
3. F	GGATGAAGTGAA-TTTTGA-----AGTATATTAA
4. F	GGATTAGGGG-----TGAAG--TAGTATTAATTAA
5. F	GGATAGGGT-AAG-----AGTAGTAG-----TAGTATTAA
6. F	GGATTTAGAG-----GTGAATTTTGAAGTAAATTAG
7. F	GGATAAGG-----GTGAAATTTGAAGTATATTAA
8. F	GGATAAGGTTAAG-AGAGAGATGAATTATGAAGTATATTAA
9. F	GGATTTGGGAAGGTTTAGAGGTGAATTTTGTAGTATATTAA
10. F	GGATGAGGGG-----TGAATTATGGAGT-TATTAA
11. F	GGATTGGGGAAGGTTTGAGATGAATTTTGAGAGTATATTAA
12. F	GGATTTGGTGAAGTA-----AA
dNTP	
13. F	GGATCCGGCCAAGCTTCAAGCTGAATTCTGCAGTACATTAA
14. R	GGATCCG----AGCGTCGAGCTGAATTCTGCAGTCCATTAT
15. F	GGATCCGGCCAAGCTTTCGAGCTGAATCCTGCAGTACATTAA
16. F	GGATCCGGCCAAGCTTCAAGCTGAATTCTGCAGTACATTAA
17. R	GGATCCGGCCAAGCTTTCGAGCTGAATTCTGCAGTACATTAG
18. R	GGATCCGGCCAAGCTTTCGAGCTGAATTCTGCAGTACATTAA
19. R	GGATCCGGCCAAGTTTCGAGCTGAATTCTGCAGTACATTAA
20. F	GGATCCGGCCAAGCTTTCGAGCTGAATTCTGCAGTACATTAA

Sloppymerase™

- what else fun can you do with it?

Expressing Sloppymerase™ in bacteria

Can you speed up evolution of recombinant proteins?

Expressing Sloppymerase™ in mamalian cells

Can you use it as a model system for cancer – increasing mutation rate in defined cell populations?



Suggested reading

Analysis of protein interactions in situ by proximity ligation assays

Koos B, Andersson L, Clausson CM, Grannas K, Klaesson A, Cane G, Söderberg O.

Curr Top Microbiol Immunol. 2014, 377:111-26

Proximity ligation assays: a recent addition to the proteomics toolbox

Weibrecht I, Leuchowius KJ, Clausson CM, Conze T, Jarvius M, Howell WM, Kamali-Moghaddam M, Söderberg O.

Expert Rev Proteomics. 2010, 7(3):401-9

Next generation Pathology – surveillance of tumor microecology

Koos B, Kamali-Moghaddam M, David L, Sobrinho-Simões M, Dimberg A, Nilsson M, Wählby C, Söderberg O.

J Mol Biol. 2015, 427(11): 2013-2022

Let there be light!

Raykova D, Koos B, Asplund A, Gelléri M, Ivarsson Y, Danielsson UH & Söderberg O

Proteomes, 2016, 4(4): 36



**TÄNKA RÄTT ÄR STORT
MEN TÄNKA FRITT ÄR STÖRRE**